



Calcitropic Principle of *Solanum glaucophyllum* in Broiler Chickens

¹S. Rovegno, ¹F. Miccoli, ^{1,3}M. Dallorso, ²B. Iglesias and ²J. Azcona

¹Facultad de Ciencias Agrarias, Universidad Nacional de Lomas de Zamora (UNLZ), Ruta 4 km 2 (1836) Llavallol, Buenos Aires, Argentina

²Estación Experimental Agropecuaria-Pergamino Instituto Nacional de Tecnología Agropecuaria (INTA), Pergamino, Provincia de Buenos Aires, Argentina

³Grupo Pecuario, Centro Atómico Ezeiza, Comisión Nacional de Energía Atómica (CNEA), Avenida del Libertador 8250, Buenos Aires, Argentina

Key words: *Solanum glaucophyllum*, poultry, broiler chickens, 1.25 dihydroxyvitamin D₃-glycoside, 1.25 dihydroxyvitamin D₃

Abstract: *Solanum glaucophyllum* (duraznillo blanco) which causes calcinosis of cattle in Argentina, contains the active metabolite of vitamin D₃ either free or conjugated with carbohydrates as 1.25-(OH)₂ Vitamin D₃-glycoside. Economic relevance of *S. glaucophyllum* could be seen as being the causal factor of toxicosis of grazing cattle and also as a valuable source of Vitamin D₃ active metabolites. Some pharmacological applications in human and veterinary medicine as well as in animal husbandry have been assayed. The calcitropic principle was thought to be the main active metabolite of vitamin D₃, once liberated from its glycoside. However, calcitropic activity of this glycoside seems to be feasible. In order to know the fate of the water soluble calcitropic principle of *Solanum glaucophyllum* administered orally to broilers, 1,25-(OH)₂ Vitamin D and 1.25-(OH)₂ Vitamin D₃-glycoside plasma levels were assayed over a period of 24 h after a unique administration of the aqueous extract of the plant. Here we show that 1.25(OH)₂ vitamin D plasma levels did not increase while 1.25 (OH)₂ Vitamin D₃-glycoside rapidly appeared in blood, remained in circulation during the 1st 6 h and was undetectable 24 h after a unique oral administration of *S. glaucophyllum* (0.11 g DM kg⁻¹ BW), to finishing broilers.

Corresponding Author:

M. Dallorso

Facultad de Ciencias Agrarias, Universidad Nacional de Lomas de Zamora (UNLZ), Ruta 4 km 2 (1836) Llavallol, Buenos Aires, Argentina

Page No.: 28-31

Volume: 8, Issue 3, 2015

ISSN: 1993-5285

Research Journal of Poultry Sciences

Copy Right: Medwell Publications

INTRODUCTION

Solanum Glaucophyllum (SG) Desfontaines (common name: duraznillo blanco) was first described in Brazil in 1846 under the name of *S. malacoxylon* Sendtner. In Argentina this species had been named

S. glaucum Dunal/Bertolini but Cabrera showed that both names were conspecific (other names: *S. glaucescens* Baile and *S. glaucumfrutescens* Larranaga). An excellent description of the growth habit, the distribution and ecology of this weed in Argentina has been already published (Okada *et al.*, 1977).

The ingestion of this plant, growing wild in South America and India (Deb, 1979), causes a calcinosis of cattle named Enteque Seco in Argentina and Uruguay and Espichamento in Brazil. The active principle associated with the pathological signs of this disease is the 1.25-(OH)₂ Vitamin D₃ (Haussler *et al.*, 1976). This metabolite of Vitamin D₃ is encountered either free or conjugated with carbohydrates as 1.25-(OH)₂ Vitamin D₃-glycoside (Weissenberg *et al.*, 1989).

Economic relevance of SG could be seen as being the causal factor of toxicosis of grazing cattle and also as a valuable source of Vitamin D₃ active metabolites. Some pharmacological applications in human and veterinary medicine as well as in animal husbandry have been assayed. Concerning human medicine, the main indications comprises chronic renal failure, hypoparathyroidism and bone disorders. Some veterinary uses have been: the prevention of milk fever, pseudo-vitamin D deficiency of pigs and acidosis in chicks (Morris, 1982; Horst *et al.*, 2003). The first animal husbandry application was the assay of the effect of SG leaves powder on egg shell quality in laying hens researchers concluded that there was a significant increase in egg shell thickness but a significant decrease in the percentage of laying. Intensive poultry production produces excess of P in manure. When this is applied on fields as fertiliser, the surplus can cause eutrophication of water bodies (Sharpley *et al.*, 1994). SG has been added to the feed of chicks with the intention to improve phosphorus utilization and so, to reduce environmental phosphorus contamination due to poultry production (Cheng *et al.*, 2004).

Additionally intensive broiler production also leads to higher occurrence of skeletal problems and this in turn, to lower performance and animal welfare (Sanotra *et al.*, 2001). A sequential study carried out on broilers suggested that the incidence of Tibial Dyschondroplasia (TD), one of the main skeletal problems was related to an inherent predisposition to rickets and to lower serum concentrations of 1.25-(OH)₂ D₃, throughout the period of maximum tibial growth (Parkinson and Thorp, 1996).

The calciotropic principle of SG was thought to be the main active metabolite of vitamin D₃, once liberated from its glycoside. The previous research done with rabbits, ewes (Dallorso *et al.*, 2000) and cows (Dallorso *et al.*, 2010) demonstrated early augmentation of 1.25-(OH)₂ Vitamin D plasma concentrations after a unique oral dose of aqueous extract of SG dry leaves. However this metabolite showed higher levels than untreated animals in plasma samples of rabbits administered with a unique subcutaneous dose of aqueous extract of SG dry leaves (Dallorso *et al.*, 2000). The biological activity of 1.25(OH)₂ Vitamin D₃-glycoside

was early demonstrated *in vitro* by Procsal *et al.* (1976) and more recently by us when we were evaluating *S. glaucophyllum* toxicity *in vivo* with rabbits.

In order to know the fate of the water soluble calciotropic principle of SG administered orally to broilers, 1.25-(OH)₂ Vitamin D and 1.25-(OH)₂ Vitamin D₃-glycoside plasma levels were assayed over a period of 24 h after a unique administration of the aqueous extract of the plant.

MATERIALS AND METHODS

***Solanum Glaucophyllum* (SG):** Leaves of the plant collected in Cañuelas in Buenos Aires province of Argentina in summer 2004 were dried, grounded and sieved 1 mm (DM) and assayed by Radioimmunoassay (RIA) to evaluate its Vitamin D Activity (VDA; [1.25D] = 10 µg g⁻¹ DM of SG) (Gil and Dallorso, 2002; Gil *et al.*, 2007). The Aqueous Extract (AE) administered to broilers was prepared as in Dallorso *et al.* (2001) (0.33 g DM mL⁻¹; render of active principle 50%) and stored at -20°C in a dark bottle closed under N₂ until its administration.

Animals and treatments: About 28 males Cobb-500, 49 days old (mean Body Weight-BW) = 3.08±0.20 kg) from Granja Tres Arroyos, Capilla del Señor, Buenos Aires, Argentina were used. Birds were assigned at random to one of three groups. Group 1 (n = 12): received a unique oral dose of 1 mL of AE of SG (0.11 g DM kg⁻¹ BW); Group 2 (n = 12): received a unique oral dose of 1 mL of run water; Group 3 (n = 4): birds of the same pen without treatment or manipulation. Animals were euthanized by complete bleeding 1 h (Group 1: n = 3; Group 2: n = 3), 3 h (Group 1: n = 3; Group 2: n = 3), 6 h (Group 1: n = 3; Group 2: n = 3) and 24 h (Group 1: n = 3; Group 2: n = 3), after each administration. Animals of Group 3 were euthanized at the beginning of the trial (time 0). Blood samples were heparinised to obtain plasma by centrifugation at 3500 rpm for 10 min, to be used for the determination of 1.25D and 1.25D₃-glycoside plasma concentrations by RIA.

Determinations: About 1 mL of plasma added with 0.050 mL ³H-1,25D₃ (1000 cpm, ~170 Ci/mM) as recovery tracer+1 mL acetonitrile were vortexed and centrifuged at 4000 rpm for 15 min. Supernatant+0.5 mL of bidistilled water were applied to solid phase extraction minicolumns Waters SPE C18 (500 mg) that have been conditioned with hexane, chloroform, methanol and bidistilled water (Dallorso *et al.*, 2001). The 1.25D₃-glycoside (F1) and 1.25D (F2) were eluted with 5 mL methanol:water (70:30) and 5 mL hexane: isopropanol (92:8), respectively. Both fractions were dried

in water bath below 40°C under N₂; F1 was resuspended in methanol: water (50:50) and F2 in ethanol, for RIA determination (Gil and Dallorso, 2002). The 1.25D₃-glycoside concentration was measured against 1.25 (OH)₂D₃ (as standard of 1.25D₃-glycoside is not available) and expressed as 1,25D₃-glycoside equivalent to 1.25D in pg mL⁻¹ plasma.

Analysis of data: About 1,25D plasma concentrations were examined by a 2-way ANOVA where one factor was TREATMENT (Grupos 1-2) and the other TIME (1- 3- 6 y 24 h). 1,25D₃-glycoside plasma concentrations (Group 1) were studied by 1-way ANOVA where the factor was TIME (1-3-6 y 24 h). Comparison of means were analysed by Tuckey. All the analysis were performed using the software Statistix SXW-Version 8.0.

RESULTS AND DISCUSSION

About 1.25D plasma levels at time 0 (Group 3) and 1-3-6 and 24 h after administrations (Groups 1 and 2) can be shown in Fig. 1. Basal mean value was 41.69±7.95 pg mL⁻¹ (n = 4). Values did not differ neither between treatments nor among time after the administration. Nevertheless, we could see the lowest mean value (27.85±4.94 pg mL⁻¹; n = 3) 3 h after the administration of SG. Here we show that 1.25 (OH)₂ Vitamin D plasma levels did not increase after the administration of a unique oral dose of 0.11 g DM kg⁻¹ BW of *S. glaucophyllum*, to finishing broilers. Other researchers have measured augmented plasma 1.25 (OH)₂ Vitamin D levels in birds treated with *S. glaucophyllum* previously incubated with β-glycosidases (Peterlik *et al.*, 1976).

About 1.25D-glycoside plasma levels at time 0 (Group 3) and 1-3-6 and 24 h after the administration of SG (Group 1) can be shown in Fig. 2. Time 0 mean value was 0 pg mL⁻¹ (n = 4). Values showed differences among time after the administration of SG by ANOVA (p = 0.05). The comparison of means showed significant differences by Tuckey (p = 0.1) of values corresponding to 1 h (18.08±4.83 pg mL⁻¹), 3 h (16.38±8.18 pg mL⁻¹) and 6 h (4.48±1.35 pg mL⁻¹) vs. those of 24 h (0 pg mL⁻¹), following the administration of SG. 1.25D-glycoside reached its peak 1 h and remained high up to 3 h, after the administration of SG while 1.25D showed its lowest plasma levels (Fig. 1 and 2). Note that the glycoside mass might be higher than expressed, here as displacement capability of 3H-1.25 (OH)₂D₃ from antibody binding sites by 1.25 (OH)₂ Vitamin D₃-glycoside has proven to be lower than that of the free metabolite of vitamin D₃ used as standard. In this research, plasma 1.25 (OH)₂ vitamin D₃-glycoside rapidly appeared in blood of broilers, remained in circulation during the 1st 6 h and was undetectable at 24 h, after a unique oral administration of *S. glaucophyllum*.

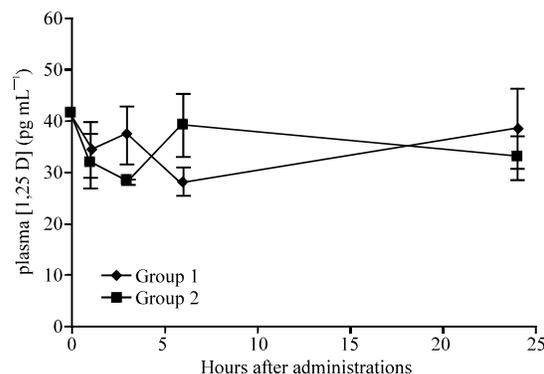


Fig. 1: 1.25 D plasma levels in male Cobb 500 finishing broilers after a unique administration of AE of SG (Group 1) or water (Group 2) by the oral route

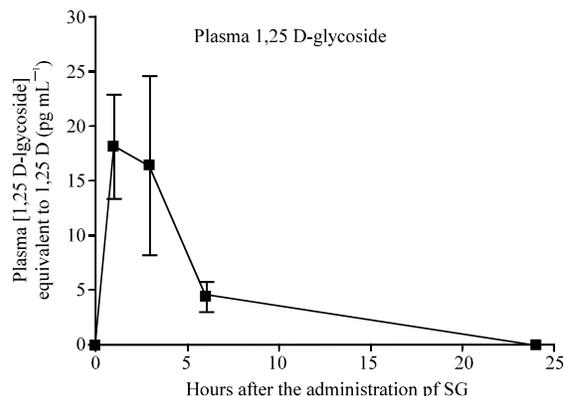


Fig. 2: 1.25 D-glycoside plasma levels in male Cobb 500 finishing broilers after a unique administration of AE of SG (Group 1) by the oral (Group 2) by the oral route

CONCLUSION

This is the 1 st time that 1.25 (OH)₂ Vitamin D and 1.25 (OH)₂ Vitamin D₃-glycoside were measured simultaneously in plasma of broiler chickens administered orally with *S. glaucophyllum*. The results obtained here are basic for those that are conducting feeding trails with *S. glaucophyllum* in poultry.

ACKNOWLEDGEMENTS

Researchers are grateful to Dr. R. Horst (USDA) for gently giving us the antibody for the analysis and to E. Rosenberg (CNEA) and M. Neme (UNLZ), for their technical assistance. The researchers also acknowledge the support provided by INTA, CNEA and UNLZ in 2009- 2010.

REFERENCES

- Cheng, Y.H., J.P. Goff, J.L. Sell, M.E. Dallorso, S. Gil, S.E. Pawlak and R.L. Horst, 2004. Utilizing *Solanum glaucophyllum* alone or with phytase to improve phosphorus utilization in broilers. *Poult. Sci.*, 83: 406-413.
- Dallorso, M., J. Azcona, B. Iglesias, S. Gil and N. Belmonte, 2010. Vitamin D Metabolism in Experimental Animals: Kinetics of the *Solanum glaucophyllum* Active Principle in Cows and Calcium, Phosphorus and Vitamin D3 Requirements in Broilers. In: Sustainable Improvement of Animal Production and Health, Odongo, N.E., M. Garcia and G. Viljoen, (Eds.), Joint FAO/ IAEA Programme, Austria, pp: 383-388.
- Dallorso, M., Pawlak, S. Gil, F. Lema and A. Marquez, 2000. 1 α ,25(OH)₂ vitamin D Plasmatic Levels in Experimental Animals Dosed with *Solanum glaucophyllum*. In: Vitamin D Endocrine System: Structural, Biological, Genetical and Clinical Aspects, Norman, A.W., R. Bouillon and M. Thomasset, (Eds.), University of California Press, USA., pp: 951-954.
- Dallorso, M., S. Gil, S. Pawlak, F. Lema and A. Marquez, 2001. 1,25(OH)₂ vitamin D concentration in the plasma of *Solanum glaucophyllum* intoxicated rabbits. *Aust. Ve. J.*, 79: 419-423.
- Deb, D.B., 1979. Solanaceae in India. In: The Biology and Taxonomy of the Solanaceae, Hawkes, J.G., R.N. Lester and A.D. Skelding (Eds.). Academic Press, London, UK., ISBN: 0123331501, pp: 87-112.
- Gil, S. and M. Dallorso, 2002. Evaluacion de la actividad vitamina D de *Solanum glaucophyllum* a traves de un radioinmunoanalisis. *Invest. Vet.*, 4: 55-61.
- Gil, S., M. Dallorso and R. Horst, 2007. Screening of vitamin D activity (VDA) of *Solanum glaucophyllum* leaves measured by radioimmunoassay (RIA). *Steroid Biochem. Mol. Biol.*, 103: 483-486.
- Haussler, M., R. Wassermann, T. Mc Cain, M. Peterlik, K. Bursac and M. Hughes, 1976. 1,25-dihydroxyvitamin D₃-glycoside: Identification of a calcinogenic principle of *Solanum malacoxylon*. *Life Sci.*, 18: 1049-1056.
- Horst, R., J. Goff, S. Gil, M. Dallorso and S. Pawlak, 2003. Using *Solanum glaucophyllum* as a source of 1,25-dihydroxyvitamin D to prevent hypocalcemia in dairy cows. *Acta Vet. Scand. Suppl.*, 98: 242-242.
- Morris, K., 1982. Plant induced calcinosis: A review. *Vet. Hum. Toxicol.*, 24: 34-48.
- Okada, K., B. Carrillo and M. Tilley, 1977. *Solanum malacoxylon* Sendtner: A toxic plant in Argentina. *Econ. Bot.*, 31: 225-236.
- Parkinson, G. and B. Thorp, 1996. Sequential studies of endochondral ossification and serum 1,25-dihydroxycholecalciferol in broiler chickens between one and 21 days of age. *Res. Vet. Sci.*, 60: 173-178.
- Peterlik, M., K. Bursac and M. Haussler, 1976. Further evidence for the 1,25-dihydroxyvitamin D-like activity of *Solanum malacoxylon*. *Biochem. Biophys. Res. Commun.*, 70: 797-804.
- Procsal, D., H. Henry, T. Hendrickson and A. Norman, 1976. 1 α ,25-dihydroxyvitamin D₃-like component present in the plant *Solanum glaucophyllum*. *Endocrinol.*, 99: 437-444.
- Sanotra, G., J. Lund, A. Ersboll, J. Petersen and K. Vestergaard, 2001. Monitoring leg problems in broilers: a survey of commercial broiler production in Denmark. *World's Poul. Sci. J.*, 57: 55-69.
- Sharpley, A., S. Chapra, R. Wedepohl, J. Sims, T. Daniel and K. Reedy, 1994. Managing agricultural phosphorus for protection of surface waters: Issues and options. *J. Environ. Qual.*, 23: 437-451.
- Weissenberg, M., A. Levy and R. Wasserman, 1989. Distribution of calcitriol activity in *Solanum glaucophyllum* plants and cell cultures. *Phytochem.*, 28: 795-798.