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Studying the Prevalence of Predominant Bacterial Micro-Organisms in MICU Along-with their Drug Sensitivity and Antimicrobial Resistance Pattern

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ABSTRACT

Incidence of different organisms in infections acquired in community as well as healthcare set ups vary across different countries and even in different studies. Emergence of new pathogenic strains as well as MDR strains has brought into focus the importance of bacterial surveillance. Further microbial flora and their susceptibility pattern to antibiotics tend to change over time in particular set up. Hence, it is necessary to have knowledge regarding spectrum of microbes and their sensitivity pattern at individual healthcare set up. The above study was conducted at the Medical intensive care unit of a tertiary care hospital from January 2012 to December 2013. Samples of the 883 patients admitted in the MICU were included in this study. All isolates were obtained from a wide range of clinical samples (e.g., urine, pus, blood, sputum, tracheal secretions). Samples were processed for culture and sensitivity testing at department of microbiology. These isolates were studied on the basis of site of infection, characteristics of patients, clinical signs and symptoms, antimicrobial resistance pattern, thus identified as true pathogens. Proportion of gram negative organisms was greater than gram positive organisms. Respiratory isolates were least resistant to Carbapenems, Piperacillin-Tazobactam. Urinary isolates were found least resistant to amikacin and meropenem. For *Klebsiella pneumoniae* resistance to meropenem increased from 27.30-50.80%. Resistance to amikacin decreased from 67.60-50%. Prevalence of bacteria differs in different health set ups. Antibiotic resistance profile for any bacterial isolate need not be similar in any two health set ups. For rational use of antibiotics, profile of prevalent organisms and their resistance/sensitivity pattern should be known.

INTRODUCTION

The fight between man and microbes has been ongoing ever since antibiotics were introduced in 20th century. Despite initial success of antibiotics in combating infections, microbes have definitely won the war. Emergence of new pathogenic strains as well as MDR strains has brought into focus the importance of bacterial surveillance. Role of this data cannot be overemphasized while choosing empirical antibiotic in patients with critical illness. Several studies have shown that outcome of patients improves with proper selection of empiric antibiotic. Incidence of different organisms in infections acquired in community as well as healthcare set ups vary across different countries and even in different studies. Further microbial flora and their susceptibility pattern to antibiotics tend to change over time in particular set up. Hence ideally it is necessary to have knowledge regarding spectrum of microbes and their sensitivity pattern in individual healthcare set up like medical intensive care unit^[1].

Therefore, the above study was conducted to find out the organisms causing infection in patients admitted in MICUs and to know the resistance pattern of isolates. Knowledge of the antibiotic susceptibility of the organisms isolated in the MICU can help to formulate an antibiotic policy for the MICU.

MATERIALS AND METHODS

Study place: The above study was conducted the above study was conducted at the medical intensive care unit of a tertiary care hospital from January 2012 to December 2013.

Study design: Observational study.

Inclusion criteria: Patients showing clinical signs of infection such as fever $>38^{\circ}\text{C}$, Leukocytosis $>10000\text{ mm}^{-3}$, infiltrates on chest x-ray, persistent tracheal aspirates/secretions, turbid urine and those who were ready to give written consent for participation were included.

Exclusion criteria: Patients having no clinical signs of infection and unwilling to give consent for participation were excluded.

Sample size: 883.

Data analysis: Data was collected in pre-defined case report format.

Ethical considerations: All the necessary permissions were taken from the institutional ethics committee before beginning the study.

All isolates were obtained from a wide range of clinical samples (e.g., urine, pus, blood, sputum, tracheal secretions) from MICU patients. Samples were

processed for culture and sensitivity testing at department of microbiology. These isolates were studied on the basis of site of infection, characteristics of patients, clinical signs and symptoms, antimicrobial resistance pattern, thus identified as true pathogens.

RESULTS

Overall in the study period, Gram negative bacilli (GNB) were most prevalent. In GNB, prevalence of *Klebsiella pneumoniae* (112/342) (33%) was highest followed by *Pseudomonas* spp. (57/342) (17%). In Gram positive organisms *Staphylococcus aureus* was predominant (66/342) (19%). Prevalence of *Klebsiella pneumoniae* was decreased from 42% in 2012 to 29% in 2013. Prevalence of *Pseudomonas* spp. increased from 14% in 2012 to 18% in 2013. Prevalence of *Staphylococcus aureus* was increased from 14% in 2012 to 22% in 2013 (Table 1 and 2).

Cefamycins. 66.70% resistance reported to Amikacin. Resistance to Ceftazidime/Clavulanic acid, Piperacillin, Meropenem and Piperacillin/Tazobactam was 71, 50, 32.40 and 31.20%, respectively (Table 3).

57.10% of respiratory isolates were sensitive to Piperacillin-Tazobactam. 54.10% sensitivity reported to Amikacin. 40.80% were sensitive to Meropenem while Fluoroquinolones were sensitive to 35.40% of respiratory isolates tested. None to 31.70% sensitivity was reported for various Cephalosporins tested (Table 4).

During the study year from 2012-2013 all urinary isolates tested for Amikacin and Meropenem were sensitive. Samples tested for Quinolones and Cephalosporins were 100% resistant. In 2013, isolates tested for Cefamycins, Cephalosporins, Quinolones were resistant. All isolates tested for Meropenem were sensitive. Sixty percent sensitivity was reported for Amikacin in 2013 as compared to 100% in 2012 (Table 5 and 6).

Table 1: Total number of isolates

| Organism | No. of isolates |
|------------------------------|-----------------|
| <i>Klebsiella pneumoniae</i> | 112 |
| <i>Pseudomonas</i> spp. | 57 |
| <i>Staph aureus</i> | 66 |
| <i>Escherichia coli</i> | 33 |
| <i>Citrobacter</i> | 20 |
| <i>Acinetobacter</i> | 14 |
| Others | 40 |
| Total | 342 |

Table 2: Numbers of isolates and organisms obtained in 2012 and 2013

| Organism | No. of Isolates in 2012 | No. of Isolates in 2013 |
|------------------------------|-------------------------|-------------------------|
| <i>Klebsiella pneumoniae</i> | 43 | 69 |
| <i>Pseudomonas</i> spp. | 15 | 42 |
| <i>Staph aureus</i> | 14 | 52 |
| <i>Escherichia coli</i> | 9 | 24 |
| <i>Acinetobacter</i> | 7 | 16 |
| <i>Citrobacter</i> | 4 | 7 |
| Others | 13 | 27 |
| Total | 105 | 237 |

Table 3: Antibiotic sensitivity pattern of respiratory isolates observed in 2012

| Antibiotic name | Antibiotic class | Antibiotic subclass | Code | No. | R (%) | I (%) | S (%) | R 95% C.I. (%) |
|-------------------------------|---------------------------|---------------------|------|-----|-------|-------|-------|----------------|
| Amikacin | Aminoglycosides | | AMK | 72 | 66.7 | 6.9 | 26.4 | 54.5-77.1 |
| Cefoxitin | Cephems | Cephameycins | FOX | 10 | 90 | 10 | 0 | 54.1-99.5 |
| Ceftazidime/clavulanic acid | Beta-lactam+Inhibitor | | CCV | 31 | 71 | 3.2 | 25.8 | 51.8-85.1 |
| Imipenem | Penems | Carbapenems | IPM | 12 | 0 | 41.7 | 58.3 | 0.0-30.1 |
| Meropenem | Penems | Carbapenems | MEM | 37 | 32.4 | 27 | 40.5 | 18.5-49.9 |
| Piperacillin | Penicillins | Ureidopenicillins | PIP | 8 | 50 | 0 | 50 | 17.4-82.6 |
| Piperacillin/tazobactam | Beta-lactam+Inhibitor | | TZP | 16 | 31.2 | 25 | 43.8 | 12.1-58.5 |
| Trimethoprim/sulfamethoxazole | Folate pathway inhibitors | | SXT | 7 | 100 | 0 | 0 | 56.1-100 |
| Cefoperazone | Cephems | Cephalosporin II | CFP | 6 | 100 | 0 | 0 | 51.7-100 |

Table 4: Antibiotic sensitivity pattern of respiratory isolates observed in 2013

| Antibiotic name | Antibiotic class | Antibiotic subclass | Code | No. | R (%) | I (%) | S (%) | R 95% C.I. (%) |
|-------------------------------|---------------------------|---------------------|------|-----|-------|-------|-------|----------------|
| Amikacin | Aminoglycosides | | AMK | 133 | 36.8 | 9 | 54.1 | 28.7-45.6 |
| Amoxicillin/clavulanic acid | Beta-lactam+Inhibitor | | AMC | 28 | 100 | 0 | 0 | 85.0-100 |
| Ampicillin | Penicillins | Aminopenicillins | AMP | 30 | 100 | 0 | 0 | 85.9-100 |
| Cefepime | Cephems | Cephalosporin IV | FEP | 32 | 75 | 3.1 | 21.9 | 56.2-87.9 |
| Cefotaxime | Cephems | Cephalosporin III | CTX | 50 | 70 | 14 | 16 | 55.2-81.7 |
| Cefoxitin | Cephems | Cephameycins | FOX | 111 | 84.7 | 3.6 | 11.7 | 76.3-90.6 |
| Ceftazidime | Cephems | Cephalosporin III | CAZ | 63 | 52.4 | 15.9 | 31.7 | 39.5-65.0 |
| Ceftazidime/clavulanic acid | Beta-lactam+Inhibitor | | CCV | 71 | 56.3 | 14.1 | 29.6 | 44.0-67.9 |
| Ceftriaxone | Cephems | Cephalosporin III | CRO | 1 | 100 | 0 | 0 | 5.5-100 |
| Ciprofloxacin | Quinolones | Fluoroquinolones | CIP | 82 | 57.3 | 7.3 | 35.4 | 45.9-68.0 |
| Clindamycin | Lincosamides | | CLI | 15 | 100 | 0 | 0 | 74.7-100 |
| Colistin | Lipopeptides | | COL | 6 | 50 | 0 | 50 | 13.9-86.1 |
| Erythromycin | Macrolides | | ERY | 31 | 96.8 | 3.2 | 0 | 81.5-99.8 |
| Gentamicin | Aminoglycosides | | GEN | 23 | 73.9 | 8.7 | 17.4 | 51.3-88.9 |
| Imipenem | Penems | Carbapenems | IPM | 1 | 100 | 0 | 0 | 5.5-100 |
| Linezolid | Oxazolidinones | | LNZ | 2 | 100 | 0 | 0 | 19.8-100 |
| Meropenem | Penems | Carbapenems | MEM | 142 | 43.7 | 15.5 | 40.8 | 35.5-52.3 |
| Piperacillin/tazobactam | Beta-lactam+Inhibitor | | TZP | 35 | 28.6 | 14.3 | 57.1 | 15.3-46.6 |
| Tetracycline | Tetracyclines | | TCY | 31 | 71 | 9.7 | 19.4 | 51.8-85.1 |
| Trimethoprim/sulfamethoxazole | Folate pathway inhibitors | | SXT | 57 | 82.5 | 3.5 | 14 | 69.7-90.9 |
| Cefoperazone | Cephems | Cephalosporin II | CFP | 97 | 77.3 | 8.2 | 14.4 | 67.5-84.9 |
| ESBL | | | ESBL | 7 | 100 | 0 | 0 | |

Table 5: Antibiotic sensitivity pattern of urinary isolates observed

| Antibiotic name | Antibiotic class | Antibiotic subclass | Code | No. | R (%) | I (%) | S (%) | R 95% C.I. (%) |
|-----------------|------------------|---------------------|------|-----|-------|-------|-------|----------------|
| Amikacin | Aminoglycosides | | AMK | 2 | 0 | 0 | 100 | 0.0-80.2 |
| Meropenem | Penems | Carbapenems | MEM | 1 | 0 | 0 | 100 | 0.0-94.5 |
| Nalidixic acid | Quinolones | Quinolones | NAL | 2 | 100 | 0 | 0 | 19.8-100 |
| Norfloxacin | Quinolones | Fluoroquinolones | NOR | 1 | 0 | 100 | 0 | 0.0-94.5 |
| Cefoperazone | Cephems | Cephalosporin II | CFP | 1 | 100 | 0 | 0 | 5.5-100 |

Table 6: Antibiotic sensitivity pattern of urinary isolates in 2012

| Antibiotic name | Antibiotic class | Antibiotic subclass | No. | R (%) | I (%) | S (%) | R 95% C.I. (%) |
|-------------------------------|---------------------------|---------------------|-----|-------|-------|-------|----------------|
| Amikacin | Aminoglycosides | | 5 | 40 | 0 | 60 | 7.3-83.0 |
| Cefotaxime | Cephems | Cephalosporin III | 2 | 100 | 0 | 0 | 19.8-100 |
| Cefoxitin | Cephems | Cephameycins | 1 | 100 | 0 | 0 | 5.5-100 |
| Ceftazidime | Cephems | Cephalosporin III | 1 | 100 | 0 | 0 | 5.5-100 |
| Ceftriaxone | Cephems | Cephalosporin III | 1 | 100 | 0 | 0 | 5.5-100 |
| Ciprofloxacin | Quinolones | Fluoroquinolones | 2 | 100 | 0 | 0 | 19.8-100 |
| Gentamicin | Aminoglycosides | | 1 | 100 | 0 | 0 | 5.5-100 |
| Meropenem | Penems | Carbapenems | 1 | 0 | 0 | 100 | 0.0-94.5 |
| Nalidixic acid | Quinolones | Quinolones | 1 | 100 | 0 | 0 | 5.5-100 |
| Nitrofurantoin | Nitrofurans | | 2 | 50 | 50 | 0 | 2.7-97.3 |
| Norfloxacin | Quinolones | Fluoroquinolones | 3 | 100 | 0 | 0 | 31.0-100 |
| Trimethoprim/sulfamethoxazole | Folate pathway inhibitors | | 4 | 100 | 0 | 0 | 39.6-100 |
| Cefoperazone | Cephems | Cephalosporin II | 2 | 50 | 0 | 50 | 2.7-97.3 |
| Trimethoprim/sulfamethoxazole | Folate pathway inhibitors | | 1 | 100 | 0 | 0 | 5.5-100 |

Table 7: Antibiotic sensitivity pattern of blood cultures in 2012

| Antibiotic name | Antibiotic class | Antibiotic subclass | No. | R (%) | I (%) | S (%) | R 95% C.I. (%) |
|-------------------------------|---------------------------|---------------------|-----|-------|-------|-------|----------------|
| Cefoxitin | Cephems | Cephameycins | 2 | 0 | 0 | 100 | 0.0-80.2 |
| Trimethoprim/sulfamethoxazole | Folate pathway inhibitors | | 2 | 0 | 0 | 100 | 0.0-80.2 |

All blood isolates were sensitive to Cephameycins and Trimethoprim/ Sulfamethoxazole in 2012 as well as in 2013. Additional 100% sensitivity was observed in 2013 to Aminopenicillins, Lincosamides and Macrolides. 66.70% sensitivity was reported to Meropenem and Piperacillin/Tazobactam. 50% resistance noted to Amikacin and Cefoperazone each. 33.33% resistance noted to Meropenem and Ceftazidime/Clavulanic acid (Table 7 and 8).

Amikacin, Piperacillin-Tazobactam and Meropenem were the most effective antibiotics against *Klebsiella pneumoniae* during study period. In 2012, 67.60% isolates of *Klebsiella pneumoniae* were resistant to Amikacin. Resistance decreased to 50% in 2013. Resistance to Meropenem increased from 27.30% in 2012 to 50.80% in 2013. Number of isolates resistant to Piperacillin-Tazobactam remained same throughout study period (50%). Resistance to

Table 8: Antibiotic sensitivity pattern of blood cultures in 2013

| Antibiotic name | Antibiotic class | Antibiotic subclass | No. | R (%) | I (%) | S (%) | R 95% C.I. (%) |
|-----------------------------|-----------------------|---------------------|-----|-------|-------|-------|----------------|
| Amikacin | Aminoglycosides | | 2 | 50 | 0 | 50 | 2.7-97.3 |
| Amoxicillin/clavulanic acid | Beta-lactam+Inhibitor | | 1 | 0 | 0 | 100 | 0.0-94.5 |
| Ampicillin | Penicillins | Aminopenicillins | 1 | 0 | 0 | 100 | 0.0-94.5 |
| Cefoxitin | Cephems | Cephameycins | 3 | 0 | 0 | 100 | 0.0-69.0 |
| Ceftazidime/clavulanic acid | Beta-lactam+Inhibitor | | 3 | 33.3 | 33.3 | 33.3 | 1.8-87.5 |
| Clindamycin | Lincosamides | | 1 | 0 | 0 | 100 | 0.0-94.5 |
| Meropenem | Penems | Carbapenems | 3 | 33.3 | 0 | 66.7 | 1.8-87.5 |
| Piperacillin/tazobactam | Beta-lactam+Inhibitor | | 3 | 0 | 33.3 | 66.7 | 0.0-69.0 |
| Tetracycline | Tetracyclines | | 1 | 0 | 0 | 100 | 0.0-94.5 |
| Cefoperazone | Cephems | Cephalosporin II | 2 | 50 | 50 | 0 | 2.7-97.3 |

Table 9: Trend in antibiotic resistance for *Klebsiella pneumoniae*

| <i>Klebsiella pneumoniae</i> | AMK | FEP | CTX | FOX | CAZ | CCV | CIP | GEN | MEM | TZP | CFP | ESBL |
|------------------------------|-------|-------|-------|-------|-------|-------|-------|-----|-------|-------|-------|------|
| 2012 | 67.60 | 25.00 | 16.66 | 16.66 | | 80.00 | | | 27.30 | 50 | 100 | |
| 2013 | 50.00 | 82.40 | 91.30 | 59.10 | 60.70 | 78.80 | 42.30 | | 50.80 | 50.00 | 95.70 | 100 |

Table 10: Trend in antibiotic resistance for *Pseudomonas aeruginosa* observed

| <i>Pseudomonas aeruginosa</i> | AMK | FEP | CTX | FOX | CAZ | CCV | CIP | COL | MEM | TZP | CFP | ESBL |
|-------------------------------|-------|-------|-------|-------|-------|-------|-------|-----|-------|-------|-------|------|
| 2012 | 60.00 | 11.11 | 44.44 | 33.33 | | 57.10 | | | 71.40 | 12.50 | 100 | |
| 2013 | 13.90 | 28.60 | 40.00 | 92.90 | 17.60 | 22.20 | 23.10 | 50 | 25.00 | 6.20 | 35.00 | 100 |

Table 11: Antibiotic resistance for *Staphylococcus aureus*

| <i>Staph aureus</i> | AMC | AMP | FOX | CIP | CLI | ERY | GEN | LNZ | TCY | SXT |
|---------------------|-----|-----|-------|-----|-----|-----|-------|-----|-------|-------|
| 2012 | | | 90.00 | | | | | | | 100 |
| 2013 | 100 | 100 | 87.20 | 100 | 100 | 100 | 69.20 | 100 | 81.00 | 83.70 |

cephalosporins increased overall during study period except for Cefoperazone (100% in 2012 and 95.70% in 2013). Resistance to Ceftriaxone and Cefoxitin increased from 16.66% in 2012 to 91.30 and 59.10% respectively. Resistance to Cefepime also increased from 25-82.40% during study period (Table 9).

Amikacin, Cefepime, Ceftriaxone, Ceftazidime, Ceftazidime/Clavulanic acid, Cefoperazone, Piperacillin-Tazobactam and Meropenem were effective antibiotics against *Pseudomonas aeruginosa* during study period. In 2012, 60% isolates of *Pseudomonas aeruginosa* were resistant to Amikacin. Resistance decreased to 13.90% in 2013. Resistance to cephalosporins was variable during study period. Resistance to Cefepime and Cefoxitin increased from 11.11% and 33.33% in 2012 to 28.60% and 92.90% in 2013 respectively. Resistance to Ceftriaxone and Ceftazidime/Clavulanic acid decreased from 44.44% and 57.10% in 2012 to 40% and 22.20% in 2013, respectively. Resistance to Ceftazidime and Ciprofloxacin was 17.60 and 23.10% in 2013. Resistance to Meropenem decreased from 71.40 to 25% during study period. Piperacillin-Tazobactam was consistently active against *Pseudomonas aeruginosa* during study period with resistance of 12.50% in 2012 and 6.20% in 2013 (Table 10).

Staphylococcus aureus isolates were totally resistant to Cefoxitin and Trimethoprim/Sulfamethoxazole in 2012. In 2013, 100% resistance was noted to Amoxicillin/Clavulanic acid, Aminopenicillins, Ciprofloxacin, Clindamycin and Linezolid. 87.20% resistance was noted to Cefoxitin. Gentamicin and Tetracycline resistance was reported to 69.20 and 81% *Staphylococcus aureus* isolates respectively (Table 11).

DISCUSSIONS

In present study bacterial isolation rate was 41.70%. In the similar study done by Radji *et al.*^[4] isolation rate was 64.68% whereas Bhaumik *et al.*^[2] from B.J. Medical College, Ahmedabad reported 39.10% isolation rate. Zaveri *et al.*^[3] from College of Medical Sciences, Amargadh, Bhavnagar reported isolation rate of 42.66%. Most common organism isolated during study period was *Klebsiella pneumoniae* (33%) followed by *Pseudomonas* spp. and *Staphylococcus aureus* (17 and 19%). Jain and Khety^[5] reported similar observation in their study conducted at Saifee Hospital, Mumbai. Bhaumik *et al.*^[2] from B.J. Medical College, Ahmedabad reported *Pseudomonas* spp. (29.12%) as most common organism isolated followed by *Klebsiella* spp. (28.08%). Zaveri *et al.*^[3] from College of Medical Sciences, Amargadh, Bhavnagar reported *E. coli*, *Acinetobacter* spp. and *Pseudomonas* spp. (21.28%). In present study, prevalence of *Klebsiella pneumoniae* decreased from 35% in 2012 to 24% in 2013. Jain and Khety^[5] reported increase in prevalence of *Klebsiella pneumoniae* in their study. Prevalence of *Pseudomonas* spp. increased from 12% in 2012 to 15% in 2013. Prevalence of *Staphylococcus aureus* increased from 14% in 2012 to 18% in 2013. Jain and Khety^[5] reported decrease in prevalence of *Staphylococcus aureus* from 25-12%. For *Pseudomonas* spp. sensitivity for Meropenem and Piperacillin-Tazobactam raised from 28.60-66.70% and from 75-93.80%, respectively, in present study. Jain and Khety^[5] noted that the sensitivity of *Pseudomonas aeruginosa* to Meropenem has decreased from 90-60%. The antibiotic that remained most active against all gram negative organisms for 2 years was Imepenem, Piperacillin-tazobactam and Amikacin.

Zaveri *et al.*^[3] also reported Piperacillin-Tazobactam being sensitive against *Pseudomonas* spp. (65%) followed by Cefoperazone-Sulbactam (55%). In present study, Amoxicillin-Clavulanic Acid found consistently resistant against *Staphylococcus aureus* over study period. Fagade *et al.*^[6] reported 42% sensitivity to Amoxicillin-Clavulanic Acid. Drug Cefoxitin and Gentamicin was found effective against *Staphylococcus aureus* (sensitivity 12.80-15.40%, respectively).

CONCLUSION

From the present study it appears that the prevalence of bacteria differs in different health set ups. Bacterial isolates seemingly prevalent in particular set up can be negligible in another setting. It is also apparent from the study that antibiotic resistance profile for any bacterial isolate need not be similar in any two health set ups. Therefore, antibiotic prescription pattern for some infection in some hospital/set up may not be applicable for another e.g., in present study sensitivity of *Pseudomonas* to Meropenem increased from 28.60-66.70% in contrast to other study which reported decrease in sensitivity from 90-60%. For rational use of antibiotics, profile of prevalent organisms and their resistance/sensitivity pattern should be known and prescription of antibiotics should be done based on local data.

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