ISSN: 1815-8846

© Medwell Journals, 2010

Does Trichoderma harzianum Really Increase Growth Parameters in Plants?

Morteza Mazhabi, Hossein Nemati, Hamid Rouhani, Ali Tehranifar,
Esmat Mahdikhani-Moghadam and Hamed Kaveh
Ferdowsi University of Mashhad, P.O. Box 91775-1163, Mashhad, Khorasan Razavi, Iran

Abstract: To study the effect of *Trichoderma harzianum* Bi on qualitative and quantitative traits of tulip and gladioluss including stem length and diameter, bud diameter, petal length, bulb perimeter and bulblet number, an experiment based on completely randomized design with 8 replications were completed in situ. Main media mixture was 40% of coco peat+40 of field soil+20% of perlite. Treatments included enriched coco peat with trichoderma in different concentrations (0, 20, 50 and 100%) of total coco peat. Results showed that 100 and 50% of enriched coco peat significantly increased stem length and diameter. These treatments also had an effect on bud diameter significantly. Petal length was also affected by 100% of enriched coco peat treatment in comparison to control. For gladiolus results showed that 50 and 100% of enriched coco peat significantly increase stem diameter in compare with 20% and control but there was not any significant difference between 100 and 50% treatments. All treatments (20, 50 and 100%) had a significant effect on bulblet appearance in both mature and immature bulbs. Mature bulb perimeter was significantly affected by all treatments, although immature bulb perimeter was only affected by 50 and 100% treatments. Flower bud formation in immature bulbs which was treated with 50 and 100% of enriched coco peat increased significantly in comparison to control. Totally, it seems that trichoderma increases tulip cut flower quality traits and induces maturity in immature bulbs. None of the treatments applied was not able to significantly change (increase or decrease) stem length, inflorescence floret number, Postharvest opened floret number and leaf length of main and lateral buds in gladiolus corms. The data obtained from the experiment show that trichoderma increases polianthes cut flowers qualitative and quantitative traits.

Key words: Culture medium, enriched coco peat, flower bud diameter, bulb quality, polianthes, Iran

INTRODUCTION

Trichoderma harzianum is a saprophytic fungus which is used generally as a biological control agent against a wide range of economically important aerial and soilborne plant pathogens (Papavizas, 1985). Trichoderma sp. have been extensively studied as potential biocontrol agents (Lynch, 1990; Papavizas, 1992). However, some studies have also shown that it can stimulate the growth of a number of vegetable and bedding plant crops (Baker, 1989; Lynch et al., 1991a, b). Lynch et al. (1991a, b) investigated the effect of Trichoderma sp. on the growth of lettuce and its ability to control damping off diseases caused by Rhizoctonia solani and Pythium ultimum. They also showed that a number of trichoderma strains had a direct effect on lettuce establishment and it is growth ratio in absence of pathogens. They found that the fungal treatments of seed reduced the emergence time of seedlings compared to the controls. From their results and those of Ousley et al. (1994), they concluded that some trichoderma strains have the potential to consistently increase plant growth (Lynch *et al.*, 1991a) and influence its phenology. Various species of trichoderma were also effective in the promotion of growth and yield in various crops (Bal and Altintas, 2006a).

T. harzianum and T. virens promoted growth of cucumber and cotton seedlings (Hanson, 2000; Poldma et al., 2000; Yedidia et al., 2001). Root and shoot growth of sweet corn was considerably increased (Bjorkman et al., 1998).

Cucumber, bell pepper and strawberry yields were increased significantly following the application of *T. harzianum* in the root zone (Poldma *et al.*, 2002; Altintas and Bal, 2005; Bal and Altintas, 2006b; Elad *et al.*, 1981). However, application of *Trichoderma* sp. was not conducive to increased yields in tomato (Bal and Altintas, 2006c). As for onion, yield and quality characteristics were not enhanced by the application of *Trichoderma* sp. (Poldma *et al.*, 2001).

Trichoderma species can improve plant growth and development (Chang et al., 1986; De Souza et al., 2008; Gravel et al., 2007; Windham et al., 1986). Growth stimulation is evidenced by increases in biomass, productivity, stress resistance and increased nutrient absorption (Hoyos-Carvajal et al., 2009).

The plant benefits by the presence of *Trichoderma* sp., suggesting an interaction as avirulent symbionts (Howell *et al.*, 2000; Harman *et al.*, 2004; Yedidia *et al.*, 1999, 2001). *Trichoderma* sp. can also produce metabolites with activities analogous to plant hormones (Cutler *et al.*, 1989, 1991).

In this study, we aimed to find if *Trichoderma* sp. had additional and promoting effects on vegetative and qualitative traits of tulip and gladiolus bulbs and corms and cut flowers.

MATERIALS AND METHODS

Inoculum preparation: *T. harzianum* Bi obtained from Ferdowsi University of Mashhad fungi collection. The isolate Bi was cultured on PDA and incubated at 25°C for 5 days. Four discs of 1.5 cm diameter were cut from the margin of trichoderma colony and added to 1 L Erlenmeyer containing 250 g. Wheat grain boiled and autoclaved for 30 min. They kept for 15 days at 25°C till covering completely the grain by trichoderma.

The grains then were mixed at the rate of 10% (v.v) with peat, autoclaved in polyethylene bags (resistant to high temp.) for 30 min and placed at 25°C±5 in laboratory condition. About 10 days later when the peat was covered by trichoderma, the contents of bags were used as trichoderma inoculums.

Culture medium: The prepared inoculums were added to the main potting mixture (40% coco peat+40% fertile soil+20% perlite) at the rate of 0 (as control), 20, 50 and 100% of used coco peat. No manure or nutritional solutions were used during this survey. Weed control and irrigation was equal in all treatments.

Bulb preparation: The 32 mature bulbs with 12±0.5 cm perimeter and 48 immature bulbs with 4±0.3 cm perimeter of tulips and 28 corms of gladiolus with 14±0.5 cm perimeter were selected.

Bulbs and corms were disinfected using sodium hypochlorite then desiccated before sowing. Pots with 20 cm diameter were used for mature and with 15 cm diameter were used for immature bulbs. The two mature and three immature bulbs were planted per pot.

Experimntal design and data analysis: To assess the effect of *Trichoderma harzianum* Bi on qualitative and quantitative traits of tulip, 2 experiments (one for tulip and one for gladiolus) were performed in situ using completely randomized design with 4 treatments and 8 replications. The data was analysed with MSTATC software. Duncan's multiple range test in p<1 and p<5% used for grouping and comparing the means.

RESULTS AND DISCUSSION

Analysis of variance for mean square of studied traits indicate that among different trichoderma concentrations applied on mature bulbs, significant differences were observed for stem length and diameter, bud diameter, bulblet number and bulb perimeter in p<1% while for petal length differences between treatments were significant in p<5% (Table 1).

Results showed that as trichoderma concentration increases, the stem length increases also. In the highest concentration of trichoderma two fold increase in comparison to control is obvious. Maximum stem length were observed in Treatment with 100% of enriched coco peat (26.46 cm) which was higher than other treatments statistically (Table 2).

Significant increase in stem diameter occurs when treatment concentration increased from 50-100%. Maximum increase in bud diameter and petal length were in 100 concentration of enriched coco peat, although there were not any significant difference between 50 and 100%

Table 1: Mean square analysis of variance for studied traits in mature tulip bulbs treated with different T. harzianum Bi concentrations

CV	df	Bulb perimeter	Bulblet number	Petal length	Bud diameter	Stem diameter	Stem length
Trichoderma	3	7.838**	9.417**	1.263*	0.332**	0.107**	323.914**
Error	28	0.850	0.431	0.316	0.022	0.009	10.412

^{*}Significant in p<5%; **Significant in p<1%

Table 2: Effect of T. harzianum Bi on studied traits in mature bulbs of tulip Darwin hybride cultivar

Eiched coco peat treatments (%)	Bulb perimeter	Bulblet number	Petal length	Bud diameter	Stem diameter	Stem length
	-		8			
100	6.125°	11.940°	5.525°	2.408°	0.775ª	26.46°
50	5.125ab	11.680°	5.137 ^{ab}	2.332ª	0.647 ^b	21.59 ^b
20	4.750 ^b	10.730 ^b	4.700^{b}	2.017 ^b	0.551°	14.89°
Control	3,500 °	9.762°	4.700 ^b	2.025 ^b	0.516°	12.50°

Means with different letter have significant difference at p<5%, Duncan's Multiple range test

treatments for these two traits (Table 2). For bulb perimeter, maximum increase in mature bulbs occur in 100% treatment while between 100 and 50% treatments differences were not significant but difference between 20 and 50% treatments was significant (Table 2).

Bulblet appearance number was in its maximum with 100% treatment. However, all treatments were significantly higher than control but there were not any significant differences between 100 and 50% treatments and 50 and 20% treatments (Table 2).

Immature bulbs also had significant differences in p<1% for bulblet number, bulb perimeter and flower bud formation in all treatments (Table 3). However, there were not any significant difference between 20, 50 and 100% of trichoderma for bulblet number but all treatment was successful to increase bulblet number significantly in comparison with control (Table 4).

Increase in *T. harzianum* Bi concentration make significant increase in immature bulb perimeter where minimum perimeter occur in control and maximum perimeter tide in 100% treatment (Table 4). Flower bud formation significantly increased when concentration of

Table 3: Analysis of variance of the effect of different inoculums percentage of *T. harzianum* Bi on immature bulbs

		Flower bud	Bulb	Bulblet		
CV	df	formation	perimeter	number		
Trichoderma	3	110.677**	3.895**	3.615**		
Error	28	27.183	0.423	0.531		

^{**}Non significant

Table 4: Effect of *T. harzianum* Bi Inoculum percent on studied traits of immature bulbs of tulip *Darwin hybride* cultivar

Eiched coco peat	Flower bud		
treatments (%)	formation	Bulb perimeter	Bulblet number
100	9.375ª	8.762ª	3.500°
50	7.813 ^{ab}	7.988 ^b	2.875ª
20	3.125^{bc}	7.438^{bc}	2.875ª
Control	1.563°	7.188°	1.875 ^b

Means with different letter have significant difference at p<5%, Duncan's multiple range test

T. harzianum Bi increased from 20-50% but no more increase were observed in bud formation when trichoderma increment resumed (Table 4).

Results also showed that 50 and 100% of enriched coco peat significantly increase stem diameter in compare with 20% and control but there was not any significant difference between 100 and 50% treatments in gladiolus (Table 5). The treatments was not able to significantly change (increase or decrease) stem length, inflorescence floret number, postharvest opened floret number and leaf length of main and lateral buds in gladiolus corms (Table 6).

In this study *T. harzianum* Bi application to culture medium was statistically effective on promotion of vegetative and qualitative traits of tulip *Darwin hybride* cultivar. Ously *et al.* (1994) delineated that some trichoderma strain had inductive effects on growth of calendula and petunia. In another study, Dubsky *et al.* (2002) declare that *T. harzianum* inoculation to peat was significantly impressive on growth and flowering of poinsettia and cyclamen.

Several mechanisms by which Trichoderma sp. influences plant development were suggested such as production of growth hormones (Windham et al., 1986), solubilization of insolubleminor nutrients in soil (Altomare et al., 1999) and increased uptake and translocation of less-available minerals (Baker, 1989; Inbar et al., 1994; Kleifield and Chet, 1992). Uptake of certain minerals such as P and N is of key importance considering their role in plant growth (Johansen, 1999; Kim et al., 1997). Promotion of growth and yield by Trichoderma sp. may also be a result of increased root area allowing the roots to explore larger volumes of soil to access nutrients and increased solubility of insoluble compounds as well as increased availability of micronutrients (Altomare et al., 1999; Yedidia et al., 2001). However, initially trichoderma must be able to establish

Table 5: Mean square analysis of variance for the studied traits in gladiolus corms treated with different T. harzianum Bi concentrations

					Number of opened	
CV	df	Stem length	Stem diameter	Floret number	florets in postharvest	Leaf length
Trichoderma	3	936.69 ^{NS}	4.149**	2.988 ^{NS}	$2.095^{\rm ns}$	9507.412 ^{NS}
Error	24	533.21	0.650	2.400	1.960	7527.160

^{**}Significant at p<1%, *Significant at p<5%, NS: Non Significant

Table 6: The effect of T. harzianum Bi on the studied traits in corms of gladiolus

Eiched coco peat			No. of opened			
treatments (%)	Stem length	Stem diameter	Floret number	florets in postharvest	Leaf length	
100	102.2ª	4.36°	12.57ª	10.29ª	279.3ª	
50	102.3a	4.22°	12.29ª	9.71°	295.7ª	
20	83.79ª	3.35 ^b	11.14ª	9.14°	327.2ª	
Control	80.86ª	2.72 ^b	12.43ª	10.29ª	362.8⁴	

Means with different letter in the same column have significant difference at p<5%, Duncan's multiple range test

an interaction with the root system. The ability of a Trichoderma species to colonize the root system of a plant depends also on the plant species.

The increased growth response of plants caused by *T. harzianum* depends on the ability of the fungus to survive and develop in the rhizosphere (Kleifield and Chet, 1992). A possible mechanism for increased plant growth is an increase in nutrient transfer from soil to root which is supported by the fact that trichoderma can colonize the interior of roots (Kleifield and Chet, 1992). Increasing effects of *T. harzianum* on plant growth and yield was suggested to be more pronounced in soils relatively poor in nutrients (Rabeendran *et al.*, 2000). Availability of water in the soil may play an important role in facilitating establishment and effectiveness of trichoderma in the soil (Altintas and Bal, 2008).

The production of plant growth hormones or analogues is another mechanism by which strains of trichoderma can enhance plant growth. About 162 species of fungi have been reported to produce auxins which are key hormones affecting plant growth and development that can be produced by fungi in both symbiotic and pathogenic interactions with plants (Gravel et al., 2007; Losane and Kumar, 1992; Patten and Glick, 2002; Shayakhmetov, 2001). In a study, seed germination of cucumber increased by application of trichoderma and this increment may be due to hormonal factors secretion like gibrellins, auxins or ethylene (Akter et al., 2007). In addition to having a stimulating effect on plant growth, exogenous IAA in the rhizosphere can also have a detrimental effect on the elongation of roots over a wide range of concentrations. Such an effect has been associated with an increase in the level of ethylene in the plant (Glick et al., 1997, 1998). IAA can increase the activity of ACC synthase which catalyses the conversion of S-adenosyl methionine to ACC, the precursor of ethylene in the plant (Kende, 1993).

The plant growth stimulation reported in Gravel et al. (2007) is most likely, the synergic result of numerous modes of action exhibited by *T. atroviride* including a regulation in the concentration of IAA in the rhizosphere and a regulation of the concentration of ethylene within the roots. The results reported by Vinale et al. (2008), clearly indicated that some trichoderma secondary metabolites are directly involved in the trichoderma-plant interactions and particularly that the compound 6PP may be considered to act as an auxin like compound and/or may act as an auxin inducer.

The identification of new molecular effectors may support the application of new biopesticides and biofertilizers based on trichoderma metabolites to be used instead of the living microbes as elicitors of plant defense mechanisms and plant growth stimulants (Vinale *et al.*, 2008). According to the results of privous studies conducted over the past 5-10 years as well as (Harman, 2006) and results of *T. harzianum* on traits studied in this experiment, it can be concluded that promotional effects of trichoderma may be due to these reasons:

- Control of root and foliar pathogens
- Induced resistance
- Biological control of diseases by direct attack of plant pathogenic fungi
- Changes in the microfloral composition on roots
- Enhanced nutrient uptake (not limited to nitrogen)
- Enhanced solubilization of soil nutrients
- Enhanced root development
- Increased root hair formation
- Deeper rooting

CONCLUSION

In this study, some Trichoderma species can be a determining factor impacting on the microbial community in the rhizosphere to enhance or even inhibit plant growth and occasionally establishing a positive interaction within plant roots as an endophyte. The latter association may be the most predictive for the selection of specific strains that can be used as bioinculants to improve crop health and productivity.

Direct plant-fungi interactions induce changes in both the fungus and plant transcriptomes affecting genes that regulate plant physiology including growth and plant defenses (Harman, 2006; Woo et al., 2006; Hoyos-Carvajal et al., 2009). In this study, the researchers found *Trichoderma harzianum* strain Bi that could stimulate both early stages of growth (flower bud formation in immature bulblets) and late stages of growth (stem, petal and mature bulb properties) in tulips but it was not significantly effective in growth stimulation of gladiolus corms. This may happen because of different reactions in different species to *T. harzianum* Bi activities or also trichoderma could have transudation of some materials in the soil which may have allelopathic effects on gladiolus corms.

REFERENCES

Akter, Z., M. Weinmann and G. Neumann, 2007.

Development of a rapid bio-test to study the activity potential of biofertilizers. Article in Proceedings, Published in: Between Tradition and Globalisation, 9th German Scientific Conference on Organic Agriculture, Hohenheim University. Stuttgart, Deutschland, 20.23.03.2007.

- Altintas, S. and U. Bal, 2005. Trichoderma harzianum application increases cucumber (*Cucumis sativus* L.) yield in unheated glasshouse. J. Applied Horticult., 7: 25-28.
- Altintas, S. and U. Bal, 2008. Effects of the commercial product based on *Trichoderma harzianum* on plant, bulb and yield characteristics of onion. Scientia Horticult., 116: 219-222.
- Altomare, C., W.A. Norvell, T. Bjorkman and G.E. Harman, 1999. Solubilization of phosphates and micronutrients by the plant-growth-promoting and biocontrol fungus *Trichoderma harzianum* rifai 1295-22. Applied Environ. Microbiol., 65: 2926-2933.
- Baker, R., 1989. Improved *Trichoderma* spp. for promoting crop productivity. Trends Biotechnol., 7: 34-38.
- Bal, U. and S. Altintas, 2006a. A positive side effect from *Trichoderma harzianum*, the biological control agent: Increased yield in vegetable crops. J. Environ. Prot. Ecol., 7: 383-387.
- Bal, U. and S. Altintas, 2006b. Application of the antagonistic fungus *Trichoderma harzianum* (TrichoFlowWPTM) to root zone increases yield of bell peppers grown in soil. Biol. Agric. Horticult., 24: 149-163.
- Bal, U. and S. Altintas, 2006c. Effects of *Trichoderma harzianum* on the yield and fruit quality of tomato plants (*Lycopersicon esculentum*) grown in an unheated greenhouse. Aust. J. Exp. Agric., 46: 131-136.
- Bjorkman, T., L.M. Blanchard and G.E. Harman, 1998. Growth enhancement of shrunken-2 sweet corn by *Trichoderma harzianum* 1295-22: Effect of environmental stress. J. Am. Soc. Horticult. Sci., 123: 35-40.
- Chang, Y.C., R. Baker, O. Kleifeld and I. Chet, 1986. Increased growth of plants in the biological control agent *Trichoderma hazianum*. Plant Dis., 70: 145-148.
- Cutler, H.G., D.S. Himmelsbach, R.F. Arrendale, P.D. Cole and R.H. Cox, 1989. Koninginin A: A novel plant growth regulator from *Trichoderma koningii*. Agricult. Biol. Chem., 53: 2605-2611.
- Cutler, H.G., D.S. Himmelsbach, B. Yagen, R.F. Arrendale, J.M. Jacyno, P.D. Cole and R.H. Cox, 1991. Koninginin B: A biologically active congener of koninginin A from *Trichoderma koningii*. J. Agricult. Food Chem., 39: 977-980.
- De Souza, J.T., B.A. Bailey, A.W.V. Pomella, E.F. Erbe, C.A. Murphy, H. Bae and P.K. Hebbar, 2008. Colonization of cacao seedlings by *Trichoderma* stromaticum, a mycoparasite of the witches broom pathogen and its influence on plant growth and resistance. Biol. Control, 46: 36-45.

- Dubsky, M., F. Sramek and M. Vosatka, 2002. Inoculation of cyclamen (*Cyclamen persicum*) and poinsettia (*Euphorbia pulcherrima*) with arbuscular mycorrhizal fungi and *Trichoderma harzianum*. Rostlinna Vyroba, 48: 63-68.
- Elad, Y., I. Chet and Y. Henis, 1981 Biological control of *Rhizoctonia solani* in strawberry fields by *Trichoderma harzianum*. Plant Soil, 60: 245-254.
- Glick, B.R., L. Changping, G. Sibdas and E.B. Dumbroff, 1997. Early development of canola seedlings in the presence of the plant growth-promoting rhizobacterium *Pseudomonas putida* GR12-2. Soil Biol. Biochem., 29: 1233-1239.
- Glick, B.R., D.M. Penrose and J. Li, 1998. A model for the lowering of plant ethylene concentrations by plant growth-promoting bacteria. J. Theor. Biol., 190: 63-68.
- Gravel, V., H. Antoun and R.J. Tweddell, 2007. Growth stimulation and fruit yield improvement of greenhouse tomato plants by inoculation with Pseudomonas putida or Trichoderma atroviride: Possible role of Indole Acetic Acid (IAA). Soil Biol. Biochem., 39: 1968-1977.
- Hanson, L.E., 2000. Reduction of Verticillium wilt symptoms in cotton following seed treatment with *Trichoderma virens*. J. Cotton Sci., 4: 224-231.
- Harman, G.E., 2006. Overview of mechanisms and uses of *Trichoderma* spp. Phytopathology, 96: 190-194.
- Harman, G.E., C.R. Howell, A. Viterbo, I. Chet and M. Lorito, 2004. *Trichoderma* species - opportunistic, avirulent plant symbionts. Nat. Rev. Microbiol., 2: 43-56.
- Howell, C.R., L.E. Hanson, R.D. Stipanovic and L.S. Puckhaber, 2000. Induction of terpenoid synthesis in cotton roots and control of *Rhizoctonia* solani by seed treatment with *Trichoderma virens*. Phytopathology, 90: 248-252.
- Hoyos-Carvajal, L., S. Ordua and J. Bissett, 2009. Growth stimulation in bean (*Phaseolus vulgaris* L.) by *Trichoderma*. Biol. Control, 51: 409-416.
- Inbar, J., M. Abramsky, D. Cohen and I. Chet, 1994. Plant growth enhancement and disease control by trichoderma harzianum in vegetable seedlings grown under commercial conditions. Eur. J. Plant Pathol., 100: 337-346.
- Johansen, A., 1999. Depletion of soil mineral N by roots of Cucumis sativus L. colonized or not by arbuscular mycorrhizal fungi. Plant Soil, 209: 119-127.
- Kende, H., 1993. Ethylene biosynthesis. Ann. Rev. Plant Physiol. Plant Mol. Biol., 44: 283-307.
- Kim, K.Y., D. Jordan and G.A. McDonald, 1997. Solubilization of hydroxyapatite by *Enterobacter agglomerans* and cloned *Escherichia coli* in culture medium. Biol. Fert. Soils, 24: 347-352.

- Kleifield, O. and I. Chet, 1992. *Trichoderma* plant interaction and its effect on increased growth response. Plant Soil, 144: 267-272.
- Losane, B.K. and P.K.R. Kumar, 1992. Fungal Plant Growth Regulators. In: Handbook of Applied Mycology: Fungal Biotechnology, Arora, D.K., K.G. Elander and K.G. Mujerji (Eds.). Marcel Dekker Inc., New York, pp. 565-602.
- Lynch, J.M., 1990. Fungi as Antagonists. In: New Directions in Biological Control: Alternatives for Suppressing Agricultural Pests and Diseases, Baker, R.R. and P.E. Dunn (Eds.). Alan R. Liss, New York, pp. 243-253.
- Lynch, J.M., K.L. Wilson, M.A. Ousley and J.M. Whipps, 1991a. Response of lettuce to *Trichoderma* treatment. Lett. Applied Microbiol., 12: 56-61.
- Lynch, J.M., R.D. Lumsden, P.T. Atkey and M.A. Ousley, 1991b. Prospects for control of *Pythium* damping-off of lettuce with *Trichoderma*, *Gliocladium* and *Enterobacter* spp. Biol. Fertil. Soils, 12: 95-99.
- Ousley, M.A., J.M. Lynch and J.M. Whipps, 1994. Potential of *Trichoderma* spp. as consistent plant growth stimulators. Biol. Fertil. Soils, 17: 85-90.
- Papavizas, G.C., 1985. *Trichoderma* and *Gliocladium*: Biology, ecology and potential for biocontrol. Ann. Rev. Phytopathol., 23: 23-54.
- Papavizas, G.C., 1992. Biological control Of Selected Soilbourne Plant Pathogens with *Trichoderma* and *Gliocladium*. In: Biological Control of Plant Diseases, Progress and Challenges for the Future, Tjamos, E.C., G.C. Papavizas and R.J. Cook (Eds.). Plenum Press, New York, pp. 223-241.
- Patten, C.L. and B.R. Glick, 2002. Role of *Pseudomonas putida* indoleacetic acid in development of the host plant root system. Applied Environ. Microbiol., 68: 3795-3801.
- Poldma, P., K. Jaakson, A. Merivee and A. Albrecht, 2000. Trichoderma viride promotes growth of cucumber plants. Proceedings of the International Conference on Development of Environmentally Friendly Plant Protection in the Baltic Region, Sept. 28-29, Tartu, Estonia, pp. 162-164.

- Poldma, P., A. Merivee, P. Johansson, J. Ascard and B. Alsanius, 2001. Influence of biological control of fungal diseases with *Trichoderma spp*. on yield and quality of onion. NJF-Seminar No. 329. New Sigths in Vegetable Production. pp 48-52.
- Poldma, P., A. Albrecht and Merivee, 2002. Influence of fungus *Trichoderma viride* on the yield of cucumber in greenhouse conditions. Proceedings of the Conference on Scientific Aspects of Organic Farming, March 21-22, Jelgava, Latvia, pp. 176-180.
- Rabeendran, N., D.J. Moot, E.E. Jones and A. Stewart, 2000. Inconsistent growth promotion of cabbage and lettuce from *Trichoderma isolates*. New Zealand Plant Prot., 53: 143-146.
- Shayakhmetov, I.F., 2001. Biological activity of metabolites from culture filtrate of *Cochliobolus sativus* and *Fusarium oxysporum* in connection with *in vitro* cellular selection of crop plants for resistance to phytopathogens. Mycol. Phytopatol., 35: 66-71.
- Vinale, F., K. Sivasithamparam, E.L. Ghisalberti, R. Marra, S.L. Woo and M. Lorito, 2008. *Trichoderma*-plant-pathogen interactions. Soil Biol. Biochem., 40: 1-10.
- Windham, M.T., Y. Elad and R. Baker, 1986. A mechanism for increased plant growth induced by *Trichoderma* spp. Phytopathology, 76: 518-521.
- Woo, S.L., F. Scala, M. Ruocco and M. Lorito, 2006. The molecular biology of the interactions between *Trichoderma* spp. phytopathogenic fungi and plants. Phytopathology, 96: 181-185.
- Yedidia, I., N. Benhamou and I. Chet, 1999. Induction of defense responses in cucumber plants (*Cucumis sativus* L.) By the biocontrol agent *trichoderma harzianum*. Applied Environ. Microb., 65: 1061-1070.
- Yedidia, I., A.K. Srivastva, Y. Kapulnik and I. Chet, 2001. Effects of *Trichoderma harzianum* on microelement concentrations and increased growth of cucumber plants. Plant Soil, 235: 235-242.