

Effect of SO₂ Treatment on Stability of Anthocyanin Pigments in Berries

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Abstract: Anthocyanins are natural pigments widely distributed in nature. Anthocyanin color molecules are a subclass of flavonoids. They are responsible for the reds, purples and blues in many flowers, fruits and vegetables. Fruits and berries are the most sample sources of anthocyanins in nature. Berries and fruits are an important part of the Finnish diet. In many researches the positive effect of fruit and berry intake on human health has been reported. Anthocyanins are considered to contribute to the healthiness of fruits and berries for their antioxidant, anti-carcinogenic, anti-inflammatory and anti-angiogenic properties for example. Anthocyanins can also improve the nutritional value of processed foods by preventing oxidation of lipids and proteins in the food products. However, the stability of anthocyanins becomes most significant also in this case, as in the case of color quality. In this study the anthocyanin pigment was extracted from the three different berries (*Morus nigra* L., *Morus alba* var. *nigra* and *Fragaria* L.). Using the soaking and wetting in ethanol (1% acidified). The extracted anthocyanin pigments then were exposed to three different concentration of SO₂ (25, 50, 100 ppm). Three groups of anthocyanin solutions keeping in darkness and refrigerator for 63 days and per 3 week, the quantity of anthocyanin absorbance recorded in 520 nm.

Key words: Anthocyanin, SO₂, degradation, berries, stability

INTRODUCTION

There is worldwide interest in using food colorants from natural sources to replace artificial colorants. Anthocyanins are a group of naturally occurring phenolic compounds, which are responsible for the attractive colors of many flowers, fruits (particularly in berries), vegetables and related products derived from them. These polyphenolic substances are glycosides of polyhydroxy and polymethoxy-derivatives of 2-phenylbenzopyrylium or flavilium salts (Wang, and Xu, 2007).

The interest in the field of anthocyanin chemistry has been generated by restriction and limitation of the use of synthetic dyes as food ingredients. Because of low toxicity of anthocyanins (Timberlake and Henry, 1986) they have a high potential as a food colorant as the substitute of synthetic red dyes. Likewise other flavonoids, the significance of anthocyanins has been discussed in relation to a wide range of physiological functions such as improvement of vision, anticancer activity and also in their implication in neural dysfunction and cognitive decline (Cantuti-Castelvetri *et al.*, 2000; Hou *et al.*, 2004; Aruoma *et al.*, 2003; Kowalczyk *et al.*, 2003).

Anthocyanins have four different structures, which are in equilibrium and include flavylum cation, quinoidal base, carbinol pseudobase and chalcon (Fig. 1). The relative amounts of these structures in equilibrium are

varied and depend on the pH and anthocyanin structure (Mazza and Minitiati, 1993). Some anthocyanins are more stable than other depends on their molecular structure.

The intensity and stability of anthocyanin pigments is dependent on various factors including structure and concentration of pigments, pH, temperature, light, intensity, quality and presence of other pigments together, metal ions, enzymes, oxygen, ascorbic acid, sugar and sugar metabolites, sulfur oxide, etc (Mazza and Minitiati, 1993; Francis, 1989).

So far, anthocyanins have not been broadly used in foods and beverages. Since they are not as stable as synthetic dyes.

Sulphur dioxide, often abbreviated to sulphite or SO₂, is used extensively in modern winemaking. It's use is predominantly for its suppression of yeast and bacterial action and its anti-oxidant properties. It is possible to make good wine without using sulphites, but these results in less control, consistency or biological stability. Sulphit is a natural by product of yeast and as much as 41 ppm has been recorded in fermentations where no SO₂ has been added (Abrahamson, Lyle, 1991).

The initial anthocyanin content and condition of processing and storage have been shown to influence the color quality of muscadin wine and similar products. (Ballinger *et al.*, 1973; Ballinger *et al.*, 1974; Bates, 1980; Flora, 1976; Flora *et al.*, 1977; Flora *et al.*, 1978; Nesbitt *et al.*, 1974).

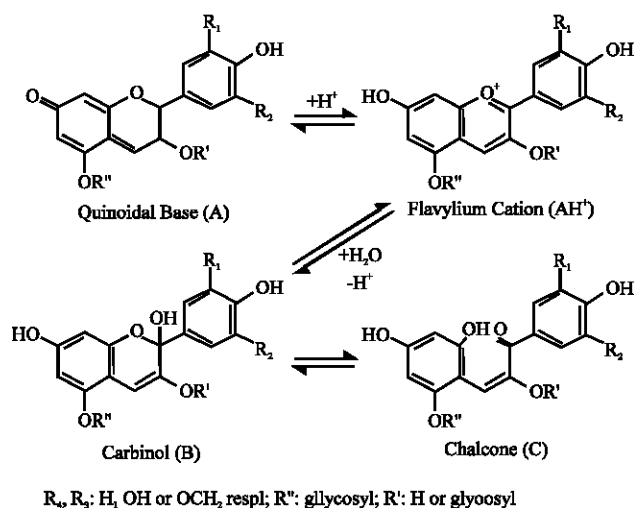


Fig. 1: Structural transformation of anthocyanins in aqueous solution

Sulfur dioxide is used widely in the wine industry as an antioxidant and inhibitor of undesirable microbial growth, but is known to bleach or lighten the color of wine due to formation of anthocyanin-4-bisulfite, a colorless compound (Burroughs *et al.*, 1975; Jurd, 1964; Robinson *et al.*, 1966; Timberlake and Bridle, 1967) Sulfur dioxide addition has a negative effect on red wine colour (Robert, 2002).

As, the usage of sulfur dioxide in wine and fruit juice industry is necessary, therefore, monitoring of suitable amount of it, is important. As, it could play its protective role and don't change the stability of color. The purpose of this study was to examine the effect of three concentration of sulfur dioxide on stability of anthocyanin pigments in berries and determine of suitable concentration.

MATERIALS AND METHODS

Sample preparation: Samples of *Berries* were obtained locally. *Berries* fruits were washed with distilled water and kept frozen at -18°C until use. The experiments were done in December to February in 2006-2007 at biochemistry lab in Urmia university.

Methods: Extraction was carried out by the methods of Chiriboga and Francis (1970). Briefly, after taking the samples out of the freezer, they left at room temperature for 30 min to defrost. Then 1000 g from each samples was put into a mixer and after adding ethanol solvent was mixed for 10 min. Then the products were filtered in Buchner funnel vacuum and Wattman filter (grade 1), the remains of the each mixture left on the filter paper was washed again with the above mentioned solvent and filtered again to get a Clear liquid. The filtered product

then placed in a balloon container within a vacuum evaporator at 35°C to separate the ethanol-acid solvent. The balloon container was separated from the vacuum evaporator and distilled water was added to dissolve the powder, which was formed at the bottom of the balloon container. The product then transferred to a 1000 mL container and brought the volume to 1000 ml using distilled water and centrifuged at 8000 rpm, the supernatant was separated and kept for further analysis.

Treatment with SO₂: To examine the effect of SO₂, three various levels of SO₂ (25, 50, 100 ppm) selected. Ninety milliliter of anthocyanin solution spilled in three groups of test tube with three repeats in each group. Certainly at first, the pH of anthocyanin solution regulated with pH meter (pH2) subsequent, different concentration of SO₂ added to anthocyanin solution. Three groups of anthocyanin solutions were kept in darkness within refrigerator for 63 days and every three weeks, the quantity of anthocyanin absorbance in 520 nm recorded.

Statistical analysis: Statistical analysis of the data was performed by ANOVA using Microsoft SAS. All experiments were repeated three times.

RESULTS AND DISCUSSION

Effect of SO₂: In blackberry (*morus nigra*), effect of time and different concentrations of SO₂ alone weren't significant, also mutual effects of time and concentration don't have significantly differences. But, totally in the case of concentration, gradually with increase in SO₂ concentration, quantities of absorbance were increased. In the case of time, lowest absorbance concerned with third time and highest was primary time.

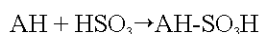
In *morus alba var nigra*. Effect of time and concentration were significant only, but their mutual effect, don't significant. In the case of time, the quantity of absorbance means from primary to third times have significant differences with zero time mean, but don't have significant differences within themselves. In the case of concentration, different concentration means have significant differences with zero concentration means. The highest absorbance related to third concentration, i. e. highest concentration of SO₂ (100 ppm) and lowest were related to zero concentration, i.e., with out SO₂.

In *Fragaria* (strawberry), effect of time was significant, but effect of concentration weren't. In the case of time, along time we observed decreased absorbance but in the case of concentration, although they haven't significant differences, gradually with increase in SO₂ concentration, observed increased absorbance of anthocyanin.

DISCUSSION

Sulfur dioxide is used widely in the wine and fruit juices as an antioxidant and inhibitor of undesirable microbial growth, but is known to "bleach" or lighten the color of wine due to the formation of anthocyanin-4-bisulfite, a colorless compound (Burroughs, 1975; Jurd, 1964; Robinson *et al.*, 1966; Timberlake, 1967).

SO₂ conjugated with anthocyanin and produce colorless product; as follow:



An equilibrium constant, for this reaction is high and expressive that little amount of SO₂ may decolorize most amounts of anthocyanins (Timberlake and Bridle, 1967). Bisulfite rapidly react with flavylum ions and if eliminated from system, reaction rapidly acts in revers direction.

Within acidified solution with pH1, the decolorization reaction is reversible. ks is obviously dependent to pH. But this is from this truth that SO₂ exists in wines balancing and when decreased equilibrium from state that react whit anthocyanin converted to state that can't react with anthocyanin demonstrated that effect of free SO₂ is much more from pH (Sapers and Simmons, 1998). Each dye that cause disrupted binary bonds in system, result in colorless. Existence of positive oxonium ions on C-2 gets anthocyanins susceptible to invasion nucleofilic compound such as SO₂ and H₂O₂. after addition SO₂, whit formation of flavon-4-sufonic acid, anthocyanins getting colorless. With deletion this material from environment in low pH condition, the anthocyanin's color is reversible. Although this decreasing in color isn't demanded in many foods, but SO₂ used along preparation process of foods (Sapers and Simmons, 1998).

The potential use of anthocyanins as food color additives has been restrained by their sensitivity to nucleophilic additions leading to their decolourization. This could occur in presence of sulfur dioxide as well, this one being widely used in the food industry as preservative. Even though this phenomenon has been observed for many years, the involved reaction still is not thoroughly understood. A former mechanism was considering the chalcone 3, resulting from the opening of carbinol2 (in equilibrium with anthocyanin 1) to be involved, leading to a chalcone-bisulfite adduct 4. More recent works ruled out this proposal and showed that the colorless compound, formed upon bisulfite addition either to C₂ or to C₄ of the anthocyanin, should be a sulfonate 5 or 6, similar to the carbinol 2 (Fig. 2).

Sims and Morris (1984) reported that Sulfur dioxide levels higher than 25 ppm free SO₂ severely bleached the color of red muscadine wine and lessened browning in high pH wine only.

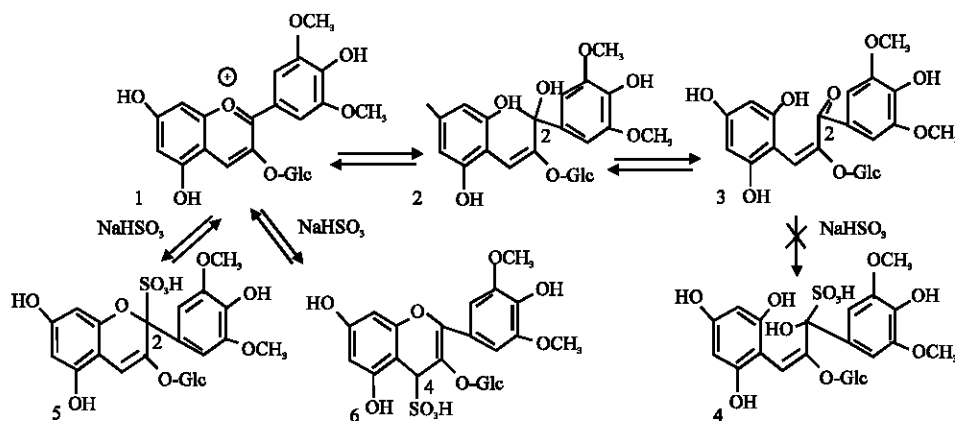


Fig. 2: Formation of colorless compound, upon bisulfite addition either to C₂ or to C₄ of the anthocyanin

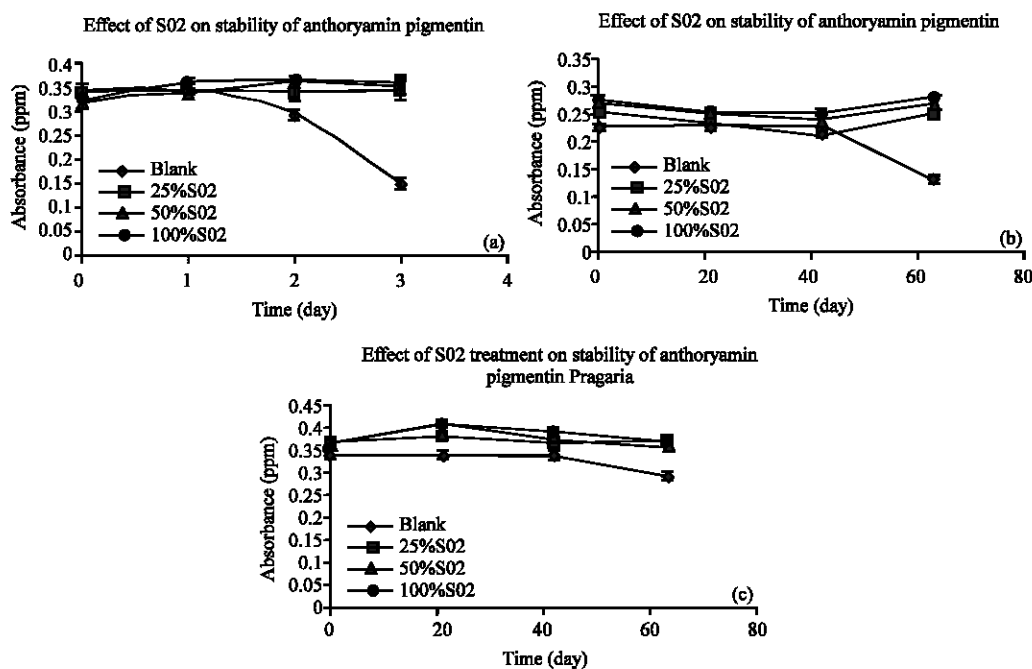


Fig. 3: Changes in anthocyanin content of enhanced juices with SO₂ during storage, at 0, 21, 42 and 63 days. Mean of 3 measurements±SE Blank, is anthocyanin solution without SO₂

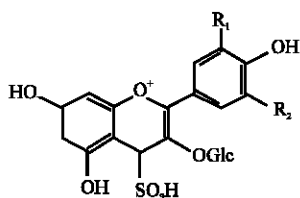


Fig. 4: Colourless bisulfite addition compound showing blocked site at C₄

The main effects of SO₂ indicate that high SO₂ levels did not inhibit browning or anthocyanin loss during storage (Fig. 3).

Robert Paul has been reported that Sulfur dioxide addition has a negative effect on red wine colour. The aromatic nature of the anthocyanin's C ring is destroyed by the reaction of the anthocyanin with sulfur dioxide to form a bisulfite addition compound. This results in a colourless molecule (Fig. 4).

REFERENCES

- Abrahamson, Lyle C., 1991. Sulfite production in wine fermentations: How much sulfite is produced by yeast in grape must with on added sulfites? Article's author is enologist at hallcrest vineyards: Organic wine works, Fenton, Santa Cruz County, California.
- Aruoma, O.I., T. Bahorun and L. Jen, 2003. Neuroprotection by bioactive components in medicinal and food plant extracts. *Mutat Res.*, 544: 203-215.
- Ballinger, W.E., E.P. Maness and W.B. Nesbitt, 1973. Anthocyanins of black grapes of 10 clones of *Vitis rotundifolia* Michx. *J. Food Sci.*, 38: 909-1000.
- Ballinger, W.E., E.P. Maness, W.B. Nesbitt, D.J. Makus and D.E. Carroll, 1974. Comparison of anthocyanins and wine color quality in black grapes of 39 clones of *Vitis rotundifolia* Michx. *J. Am. Soc. Hortic. Sci.*, 99: 338-341.
- Bassa, I.A. and F.G. Francis, 1987. Stability of anthocyanins from sweet potatoes in a model beverage. *Aresearch not. J. Food Sci.*, 52: 1753-1754.
- Bates, R.P., D. Mills, J.A. Mortensen and J.A. Cornell, 1980. Prefermentation treatments affecting the quality of muscadine grape wine. *Am. J. Enol. Vitic.*, 31: 136-142.
- Bridle, P. and C.F. Timberlake, 1997. Anthocyanins as natural food colours-selected aspects. *Food Chem.*, 58: 103-109.
- Burroughs, L.F., 1975. Determining free sulfur dioxide in red wine. *Am. J. Enol. Vitic.*, 26: 25-29.
- Cantuti-Castelvetri, I., B. Shukitt-Hale and J.A. Joseph, 2000. Neurobehavioral aspects of antioxidants in aging. *Int. J. Dev. Neurosci.*, 18: 367-381.

- Chiriboga, C. and F.J. Francis, 1970. An anthocyanin recovery system from cranberry pomace. J. Am. Soc. Hort. Sci., 9: 223.
- Flora, L.F., 1976. Time-temperature influence on muscadine grape juice quality. J. Food Sci., 41: 1312-1315.
- Flora, L.F., 1977. Storage stability of juices and jellies made from muscadine grapes (*Vitis rotundifolia* Michx.). Am. J. Enol. Vitic., 28: 171-175.
- Flora, L.F., 1978. Influence of heat, cultivar and maturity on the anthocyanin-3, 5-diglucosides of muscadine grapes. J. Food Sci., 43: 1819-1821.
- Francis, F., 1989. Food colorants: Anthocyanins. Critical Reviews in Food Science and Nutrition, 28: 273-314.
- Francis, F., 1989. Food colorants: Anthocyanins. Critical Reviews in Food Science and Nutrition, 28: 273-314.
- Harborne, J.B., 1967. Comparative Biochemistry of the Flavonoids, Academic Press, New York and London.
- Hou, D., K. Kai, J. Li, S. Lin, N. Terahara and M. Wakamatsu *et al.*, 2004. Anthocyanidins inhibit activator protein 1 activity and cell transformation: Structure activity relationship and molecular mechanisms. Carcinogenesis, 25: 9-36.
- Igarashi, K. and K. Inagaki, 1991. Effects of the major anthocyanin of wild grape (*Vitis coignetiae*) on serum lipid levels in rats. Agric. Biol. Chem., 55: 285-287.
- Igarashi, K., S. Abe and J. Sato, 1990. Effects of Atsumi-Kabu (red turnip, *Brassicacampestris* L.) anthocyanin on serum cholesterol levels in cholesterol-fed rats. Agric. Biol. Chem., 54: 171-175.
- Jurd, L., 1964. Reactions involved in sulfite bleaching of anthocyanins. J. Food Sci., 29: 16-19.
- Kamei, H., T. Kojima, M. Hasegawa, T. Koide, T. Umeda, T. Yukawa and K. Terabe, 1995. Suppression of tumor cell growth by anthocyanins *in vitro*. Cancer Invest., 13: 590-594.
- Kowalczyk, E., P. Krzesiński, M. Kura, B. Szmigiel and J. Aszczyk, 2003. Anthocyanins in medicine. Pol. J. Pharmacol., 55: 699-702.
- Markakis, P., 1982. Anthocyanins as Food Colors, Academic Press, New York.
- Mazza, G. and E. Minitati, 1993. Introduction. In Anthocyanin in Fruits, vegetables and grains, Boca raton, FL: CRC Press (Chapter 1), pp: 1-28.
- Nakamura, M., Y. Takeuchi, K. Miyanaga, M. Seki and S. Furusaki, 1999. High anthocyanin accumulation in the dark by strawberry (*Fragaria ananassa*) callus. Biotechnol. Lett., 21: 695-699.
- Nesbitt, W.B., E.P. Maness, W.E. Ballinger and D.E. Carroll, Jr. 1974. Relationship of anthocyanins of black muscadine grapes (*Vitis rotundifolia* Michx.) to wine color. Am. J. Enol. Vitic., 25: 30-32.
- aul, R., 2002. Micro-Oxygenation-Where Now? Australian Society of Viticulture and Oenology Use of Gases in Winemaking Seminar, Wine Network Australia Pty Ltd.
- Robinson, W.B., J.J. Bertino and J.E. Whitcombe, 1966. Objective measurement and specification of color in red wines. Am. J. Enol. Vitic., 17: 118-125.
- Sapers, G.M. and G.F. Simmons, 1998. Hydrogen peroxide disinfection of minimally processed fruits and vegetables. Food Tech., 52: 48-52.
- Sims, C.A. and J.R. Morris, 1984. Effects of pH, sulfur dioxide, storage time and temperature on the color and stability of red muscadine grape wine. Am. J. Enol. Vitic., 35: 1:35-39.
- Timberlake, C.F. and B.S. Henry, 1986. Plant pigments as natural food colors. Endeavour (OXF), 10: 31-36.
- Timberlake, C.F. and P. Bridle, 1967. Flavylum salts, anthocyanidins and anthocyanins. II. Reactions with sulfur dioxide. J. Sci. Food Agric., 18: 479-485.
- Wang, W.D. and S.Y. Xu, 2007. Degradation kinetics of anthocyanins in blackberry juice and concentrate. J. Food Eng., 82: 271-275.