

Comparison of Bactericidal and Fungicidal Activities of Cu (II) and Ni (II) Complexes of *Para*-Methoxy and *Para*-Hydroxy Benzoic Acid Hydrazide

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Abstract: Bactericidal and fungicidal were carried out on Cu(II) and Ni(II) *paramethoxy* and *parahydroxy* benzoic acid hydrazides activities of the synthesized compounds were screened against 12 microorganisms viz: *Yersinia* sp., *Klebsilla* sp., *Saccharomyces cerevisiae*, *Candida albicans*, *Rhodospirium* sp., *Proteus vulgaris*, *Staphylococcus aureus* and *E. coli*. The biological properties of the metal complexes revealed that in general complexation led to enhanced activity. The Cu(II) complexes appears to be much more potent antibacterial agents than the Ni(II) complexes *in vitro*; the three fungi used in this research *Rhodospirium* sp., *Saccharomyces cerevisiae* and *Candida* sp. are completely resistant to all the copper complexes. They are susceptible to at least one of the nickel complexes. *E. coli* and *P. vulgaris* are completely resistant to the nickel complexes.

Key words: Hydrazide, bacteria and fungi, complexes, biological properties, anti bacterial agents

INTRODUCTION

The chemistry of hydrazides has been intensively investigated in recent years. The reasons are manifold; First is the coordinating ability of these compounds to chelate metal ions particularly transition and lanthanide ions. (Fox and Gibas, 1953) A considerable number of hydrazides have been reported to demonstrate tuberculostatic antibacterial and antifungal activities (Gursoy *et al.*, 1997; Dodoff *et al.*, 1995; Tabakova and Dodoff, 1995; Rando *et al.*, 2002; Mamolo *et al.*, 2001; Rollas *et al.*, 2002). Application of Benzoic acid-[(5-Nitrothiophenyl-1)-methylene]-hydrazide on *Mycobacterium tuberculosis in vitro* has indicated a new lead for potential antituberculosis activity (Dodoff *et al.*, 1995). Isonicotinic acid hydrazide is highly selective for *Mycobacterium tuberculosis*.

In vitro studies on the biological activities of cyanoaliphatic acid hydrazides and their derivatives showed that these are active against *Tubercle bacilli* (Fox, 1952). Extension of the-CN group by methylene group retains appreciably the antibacterial activity of cyanoacetic acid hydrazide while modification of the cyano group into carboxy, amidino ammomethyl and other easily derivable groups reduces the antibacterial activity of cyanoaliphatic hydrazides. Dibasic acid hydrazides e.g., monoalkyl succinic acid dihydrazide inhibit the growth of *tubercle bacilli* in 200 μcm^{-3} concentration and there is

an appreciable difference in action due to difference in alkyl chain (Buu-Hoi *et al.*, 1953). In our earlier research we reported the synthesis and biological activities of some transition metal complexes of dithiocarbamate ion (Adeoye *et al.*, 2005).

We report here the synthesis of various Cu(II) and Ni(II) complexes of *p*-methoxy and *p*-hydroxybenzoic acid hydrazides and their bactericidal and fungicidal activities.

MATERIALS AND METHODS

Reagents and solvents: *p*-Methoxybenzoic, *p*-Hydroxybenzoic, Hydrazine hydrate and Ethanol were purchased from Aldrich. All the chemicals and solvents used were of reagent grade or purer and were used without further purification.

Synthesis of the ligands and complexes: The ligands and the complexes were synthesized according to the literature method (Odunola *et al.*, 2002, 2003).

Preparation of *p*-methoxy benzoic acid hydrazide PMBAH: *p*-methoxybenzoic acid (0.1314 moles) was added to 50 mL of ethanol and 2 mL of H_2SO_4 and then refluxed for 5 h. The excess ethanol was distilled off and the solution was left to cool. The ester was separated from the aqueous layer by adding 250 mL of water in a separating funnel.

Hydrazine hydrate (6.373 mL) was then added to the ester in 50 mL of ethanol and refluxed for 6 h. The ethanol, water and excess hydrazine hydrate were removed leaving the residual solid, (Yield 18.0 g).

Preparation of *p*-hydroxy benzoic acid hydrazide PHBAH:

0.2713 moles of *p*-hydroxybenzoic acid was added to 80 mL of ethanol with 2 mL of H_2SO_4 and then refluxed for 6 h. The excess ethanol was distilled off and the solution was left to cool. The ester was separated from the aqueous solution as before. 10.54 mL of hydrazine hydrate was added to the ester in 60 mL of ethanol and was refluxed for 6 h. The solid was recrystallised from excess ethanolic solution of *p*-hydroxybenzoic acid hydrazide.

Preparation of $\text{Cu}[\text{MBAH}]_2\text{Cl}_2$: $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ (1.30 g, 7.6 mmole) dissolved in 10 mL of 50% methanol was added in drops to 20 mL hot methanolic solution of *p*-methoxybenzoic acid (2.25 g, 15 mmole) in a 100 mL beaker while stirring at room temperature. After stirring for 1 h, the green precipitate formed was filtered under suction, washed with water and methanol and dried in vacuum over anhydrous calcium chloride. Copper (II) sulfate and nitrate and acetate derivatives of *p*-methoxybenzoic acid and *p*-hydroxybenzoic acid hydrazides were similarly prepared.

Physical measurements: The infrared spectra of the synthesized ligands and complexes were recorded on Perkin-Elmer 1000 spectrometer between $350\text{--}4000\text{ cm}^{-1}$ using KBr discs. Melting point of the compounds were determined using a Buchi (B-540) melting point apparatus.

ANTIMICROBIAL ACTIVITY ASSAY

The antimicrobial activity of the synthesized complexes was assayed using twelve microorganisms collected from the Biology Laboratory, Ladoké Akintola University of Technology, Ogbomoso, Nigeria using the disc diffusion method. The organisms are *Yersinia* sp. *Streptococcus pyogenes*, *Serratia marcescens*, *Bacillus subtilis*, *Bacillus licheniformis*, *Klebsiella* sp. *Saccharomyces cerevisiae*, *Candida* sp. *Rhodospirum* sp. *Proteus vulgaris*, *Staphylococcus aureus* and *E. coli*.

Eighteen-hour-old broth culture of the test organisms was inoculated onto the surface of already prepared nutrient agar and potato dextrose agar plates using sterile swab sticks. Sterilized paper discs (5 mm diameter) were then impregnated with stock solutions (0.1 mg mL^{-1}) of the test compounds. The discs were then placed equidistant apart on the surface of the previously inoculated plates. These were then incubated overnight at 37°C after which the zones of inhibition were measured.

QUALITATIVE ANTIMICROBIAL ASSAY

Two organisms were selected for the assay of bactericidal and fungicidal activity of the complexes. *S. marcescens* was chosen for the bactericidal activity assay because it was most sensitive to the $\text{Cu}(\text{II})$ complexes, while *Rhodospirum* sp. was selected for the fungicidal activity assay because the action of $\text{Ni}(\text{NO}_3)_2$ -*p*-MBAH against it was sustained for more than 3 days. One milliliter of the stock solutions (0.1 mg mL^{-1}) of CuCl_2 -*p*-HBAH and $\text{Ni}(\text{NO}_3)_2$ -*p*-MBAH were mixed with 9 mL of a broth culture of the test organisms in screw cap test tubes. The broth cultures were previously prepared by inoculating 9.8 mL of sterile broth with 0.2 mL of an 18 h old broth culture of the test organism. The whole solution

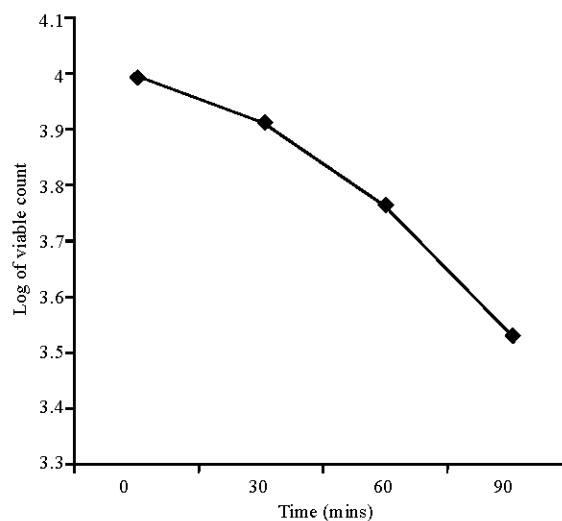


Fig. 1: Bactericidal activity of the $\text{Cu}(\text{II})$ complex against *S. marcescens*

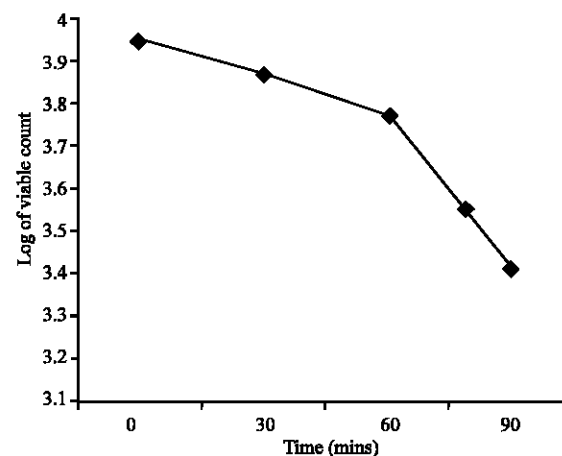
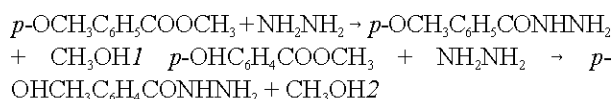


Fig. 2: Fungicidal activity of the $\text{Ni}(\text{II})$ complex against *Rhodospirum* sp.

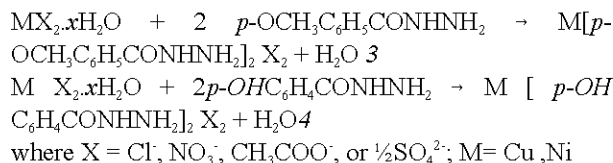
was thoroughly mixed by inverting the test tubes rapidly several times after which 0.5 mL was withdrawn aseptically at 30 min intervals, serially diluted and 0.5 mL of appropriate dilutions plated in duplicates on already prepared agar plates. The plates were then incubated overnight at 37°C and the number of colony forming units estimated and reported as the bactericidal activity (Fig. 1 and 2).

RESULTS AND DISCUSSION

The colour analytical data percentage yield elemental analysis and are given in Table 1. The *paramethoxy* and *parahydroxy* benzoic acid hydrazides were obtained from the reaction of their respective *paramethoxy* and *parahydroxy* benzoate esters and anhydrous hydrazine in fairly good yield (58-64 %) according to Eq. 1 and 2.



The ligands and the complexes were obtained in reasonable yields, the formation of the Cu(II) and Ni(II) *paramethoxy* and *parahydroxy* benzoic acid hydrazides can be represented according to the following Eq. 3 and 4.



Infrared spectra: The IR spectra bands of the ligands and complexes of the Cu(II) and Ni(II) salts are summarized in Table 2. The spectra obtained are consistent with the structural characteristics of hydrazides. The spectra also revealed that two out of the possible three binding sites are actually used in coordination. These are the 'amide I' carbonyl stretching mode $\nu_s(\text{C}=\text{O})$; which occurred in the ligands at 1600 cm⁻¹ but appeared in lower regions (1601-1604 cm⁻¹) in the complexes suggesting the coordination of the carbonyl oxygen to the metal. The 'amide II' bands consists of three different bands which may appear as distinct bands and in some cases coupling may occur between the in-plane bending $\delta(\text{N-H})$ and $\nu(\text{C-N})$ stretching mode and the stretching frequency for the amino $\nu(\text{NH}_2)$. In the ligands, the amino stretching vibrations $\nu(\text{NH}_2)$ occurs between 3395cm⁻¹ and this band becomes weaker and lowered to region between 3120-3485 cm⁻¹ in the complexes. This shift indicates that the-NH₂ group is involved in coordination to the metal. The amide II bands in the ligands and complexes occurring in the region 1534-1570 cm⁻¹ remain relatively unchanged and we can safely conclude that the $\nu(\text{C-N})_{\text{enol}}$ and $\nu(\text{C-N})_{\text{keto}}$ forms are not involved as a binding site in coordination to the metal. This has been noted in previous studies on similar systems (Rollas *et al.*, 2002; Fox, 1952).

Bioactivity of the compounds: Preliminary screening for antimicrobial activities of the stock solutions of the complexes in DMSO (100 µL) were performed using disc diffusion assay.

Overall, the Cu(II) complexes appears to be much more potent antibacterial agents than the Ni(II) complexes *in vitro* (Table 3 and 4). While the 3 fungi used in this

Table 1: Analytical data of the complexes

Compound	F.W	Colour	M.Pt °C	%M expected	%M observed
Cu[p-MBAH] ₂ SO ₄	523.85	Lt green	148 ^d	12.13	12.18
Cu[p-MBAH] ₂ Cl ₂	498.78	Ash green	158 ^d	12.74	13.04
Cu[p-MBAH] ₂ (CH ₃ COO) ₂	545.87	Deep green	150 ^d	11.64	11.94
Cu[p-MBAH] ₂ NO ₃	489.86	Bluish-green	174 ^d	12.97	12.95
Cu[p-HBAH] ₂ SO ₄	495.81	Blue	100	12.82	12.70
Cu[p-HBAH] ₂ Cl ₂	470.73	Blue	110	13.5	13.2
Cu[p-HBAH] ₂ (CH ₃ COO) ₂	517.89	Brownish-green	98	12.27	12.16
Cu[p-HBAH] ₂ NO ₃	461.81	Green	86	13.76	13.70
NiMBAH ₂ SO ₄	519.01	Blue	138	11.3	11.05
Ni[p-MBAH] ₂ Cl ₂	493.94	Green	110	11.88	12.10
Ni[p-MBAH] ₂ (CH ₃ COO) ₂	541.10	Purple	190	10.85	11.00
Ni [p-MBAH] ₂ NO ₃	485.02	Purple	170	12.10	12.00

Table 2: Relevant Infrared spectra bands

[Cu MBAH] ₂ 2Cl ₂	3345s	3058s	1602s	1534m,1517m	1300w	549vs 504s
Cu[MBAH] ₂ 2(NO ₃) ₂	3163s	3064s	1600s	1577vs1516s	1312m	699s 616s
Cu[MBAH] ₂ 2SO ₄	3261s	3055s	1600s	1570vs	1320m	550s 520s
Cu[MBAH] ₂ 2(CH ₃ COO) ₂	3155s		1600s	1542vs	1340s	567s
Ni[MBAH] ₂ 2Cl ₂	3120s	2984s	1605s	1578vs	1305s	616s
Ni[MBAH] ₂ 2(NO ₃) ₂	3256s	2901s	1601s	1578vs	1290s	617s
Ni[MBAH] ₂ 2SO ₄	3340s	3050s	1602m	1550s	1360s	599s
Ni[MBAH] ₂ 2(CH ₃ COO) ₂	3489s	3057s	1604s	1540s	1350	544s

Table 3: Response of the test organisms to the Cu (ii) complexes

Organisms	Zones of inhibition (mm)							
	CuSO ₄ <i>p</i> -HBAH	CuSO ₄ .5H ₂ O <i>p</i> -MBAH	Cu(NO ₃) ₂ <i>p</i> -HBAH	Cu(NO ₃) ₂ .H ₂ O <i>p</i> -MBAH	CuCl ₂ .H ₂ O <i>p</i> -MBAH	CuCl ₂ <i>p</i> -HBAH	Cu(OAC) ₂ <i>p</i> -HBAH	Cu(OAC).H ₂ O <i>p</i> -MBAH
<i>P. vulgaris</i>	R	10	11	12	11	R	16	R
<i>E. coli</i>	17	12	15	10	16	11	15	11
<i>Candida</i> sp.	R	R	R	R	R	R	R	R
<i>Klebsiella</i> sp.	R	R	R	R	R	13	11	R
<i>S. pyogenes</i>	16	15	R	12	R	R	R	R
<i>S. marcescens</i>	15	R	11	R	13	25	13	R
<i>B. licheniformis</i>	12	R	15	R	R	11	R	R
<i>Yersinia</i> sp.	13	10	R	17	R	10	10	12
<i>Rhodospirum</i> sp.	R	R	R	R	R	R	R	R
<i>S. cerevisiae</i>	R	R	R	R	R	R	R	R
<i>S. aureus</i>	12	R	13	R	12	10	R	R
<i>B. subtilis</i>	10	13	15	11	11	13	10	R

Table 4: Response of the test organisms to the ligands and Ni (ii) complexes

Organisms	Zones of inhibition (mm)					
	<i>p</i> -HBAH	<i>p</i> -MBAH	NiSO ₄ <i>p</i> -MBAH	NiCl ₂ <i>p</i> -MBAH	NiNO ₃ <i>p</i> -MBAH	Ni(OAC) ₂ <i>p</i> -MBAH
<i>P. vulgaris</i>	R	14	R	R	R	R
<i>E. coli</i>	15	R	R	R	R	R
<i>Candida</i> sp.	R	R	R	R	R	18
<i>Klebsiella</i> sp.	R	R	R	10	10	R
<i>S. pyogenes</i>	16	15	R	R	15	17
<i>S. marcescens</i>	12	16	R	R	11	11
<i>B. licheniformis</i>	11	R	R	R	R	16
<i>Yersinia</i> sp.	12	14	15	20	10	R
<i>Rhodospirum</i> sp.	R	R	15	R	19	20
<i>S. cerevisiae</i>	R	R	R	R	14	R
<i>S. aureus</i>	12	11	R	17	R	12
<i>B. subtilis</i>	15	13	15	R	14	13

research, *Rhodospirum* sp., *S. cerevisiae* and *Candida* sp. are completely resistant to all the Cu complexes, they are susceptible to at least one of the Ni complexes. The two organisms, *E. coli* and *P. vulgaris* completely resistant to the Ni complexes are bacterial species. The largest zone of inhibition of 25 mm was recorded against *S. marcescens* followed by 20 mm each recorded against *Yersinia* sp. and *Rhodospirum* sp. The activity of Ni(NO₃)₂ *p*-MBAH against *Rhodospirum* sp. was sustained for the longest period of over 72 h after exposure this is the reason why it was selected as the test agent for fungicidal activity. *E. coli* appears to be the most susceptible to the Cu complexes being sensitive to all the complexes tested followed by *B. subtilis*, which is sensitive to seven of the eight complexes tested. Among the bacterial species, the resistance of *Klebsiella* sp. and *S. pyogenes* to the Cu complexes are remarkable with resistance to six and seven of the tested complexes, respectively. However, the Ni complexes appears to be more potent antifungal agents, as the highest zone of inhibition of 20 mm was recorded against *Rhodospirum* while *S. cerevisiae* and *Candida* sp. are both sensitive to Ni(NO₃)₂ *p*-MBAH and Ni(OAC)₂ *p*-MBAH with zones of inhibition of 14mm and 18mm respectively. This weak antibacterial activity of the Ni

complexes is consistent with observations of Adeoye *et al.* (2005) who discovered that Ni complexes have the weakest activity when the biological activity of dithiocarbamate complexes of Zn, Pb and Ni were tested against seven bacterial isolates.

Results of the bactericidal and fungicidal activity shows a progressive decrease in the population of cells in the test medium with time for the 2 organisms tested (Fig. 1 and 2). There was a 65.5% reduction in the population of *S. marcescens* after 1 h 30 min exposure to CuCl₂ *p*-HBAH, while there was a reduction of 74.7% in the population of *Rhodospirum* sp. after the same period of exposure to Ni(NO₃)₂ *p*-MBAH. There was also a reduction in the colony size of the test organisms with time, this could be as a result of protein leakage from the cells or the physiological stress imposed on growth by exposure to the metal complexes. Microorganisms have been known to exhibit abnormal growth behaviour under adverse growth conditions.

CONCLUSION

The results obtained in this research indicates that the complexes investigated possess bactericidal and fungicidal activity.

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